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# VIRGINIA CHESAPEAKE BAY PROGRAM PHYTOPLANKTON AND PICOPLANKTON MONITORING SURVEY DATA DICTIONARY

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Virginia Chesapeake Bay Water Quality Monitoring Program: Phytoplankton and Picoplankton Components  
- Taxonomic Data Dictionary  
- Event Data Dictionary

**NOTES:**

1) THIS DICTIONARY WAS REVISED ON 3 January 2011 AND SUPERSEDES ALL OTHER DICTIONARIES FOR THE VIRGINIA PHYTOPLANKTON DATA

The Commonwealth of Virginia, in cooperation with the US EPA Chesapeake Bay Program, has monitored plankton species abundance and composition in the Virginia Chesapeake Bay mainstem and tributaries since 1985. The current program is designed to give comprehensive spatial and temporal information on phytoplankton. Sampling is performed in conjunction with the water quality monitoring programs.

**# NAMES AND DESCRIPTIONS OF ASSOCIATED DATA DICTIONARY FILE**

2000 User's Guide to Chesapeake Bay Program Biological and Living Resources Monitoring Data

**# PROJECT TITLE:**

Virginia Chesapeake Bay Monitoring Program: Lower Chesapeake Bay Phytoplankton and Picoplankton Studies

**# CURRENT PRINCIPAL INVESTIGATORS:**

>Program Manager: Frederick Hoffman, Virginia Department of Environmental Quality  
>Principal Investigator: Harold Marshall, Old Dominion University  
>Statistician: Michael Lane, Old Dominion University  
>Data Coordinator: TBD, Old Dominion University, Applied Marine Research Laboratory

**# CURRENT FUNDING AGENCIES:**

Not Applicable

**# PROJECT COST**

Not Applicable

**# QA/QC OFFICER: Not Applicable**

**# POINT OF CONTACT:**

Jacqueline Johnson  
Living Resources Data Manager  
US EPA Chesapeake Bay Program Office  
410 Severn Avenue, Suite 109  
Annapolis, MD 21403  
1-800-968-7229 x729  
1-410-267-5729  
E-Mail JJOHNSON@CHESAPEAKEBAY.NET

# LOCATION OF STUDY

Chesapeake Bay and its Tidal Tributaries in the Commonwealth of Virginia

# DATE INTERVALS

08/01/1985- 09/30/2010

# ABSTRACT

The phytoplankton monitoring program was established to determine the composition, distribution, biomass, and abundance of these plankton constituents in the Chesapeake Bay and Virginia tributaries, in relation to water quality conditions and algal productivity in these waters. Additional emphasis has been placed on bloom and toxin producing species and long term trends in population development and environmental relationships. Composite plankton collections, above and below the pycnocline, began at 7 Bay stations in July 1985. River station collections in the James, York, Rappahannock, and Pamunkey Rivers began in March 1986. Twice monthly collections were taken March through October, and once monthly November through February at all stations. Two stations were added in the Elizabeth River in February 1989, with station SBE2 discontinued in January 1998. Autotrophic picoplankton collections were added at all stations in January 1989, with plankton productivity (C-14) determinations for all stations added in July 1989. Beginning in October 1990 monthly collections were taken at all stations and this pattern continues for the Bay stations, however in 2002 collections in the tributaries for November through February were discontinued, with monthly collections taken only from March through October for the tributaries. Other modifications include discontinuing and replacing in March 1986 station RET4.1 with station TF4.2 in the Pamunkey River, and a station designation and site change in August 2005 for LE5.5 to LE5.5-W. Sampling was reduced in frequency in 2010 due to budget constraints. All below pycnocline sampling in the tributary stations was dropped in 2010.

# STATION NAMES AND DESCRIPTIONS

CB6.1 Main Channel, Mid-Bay  
 CB6.4 Main Channel, Mid-Bay  
 CB7.3E Eastern Shore Channel, Southern End  
 CB7.4 Baltimore Channel, Bay Mouth  
 LE3.6 Off Mouth of Rappahannock River  
 LE5.5 Off Mouth of James River  
 LE5.5-W Off Mouth of James River  
 RET3.1 Rappahannock River, N Buoy R10  
 RET4.1 York River,  
 RET4.3 York River, Buoy C57  
 RET5.2 James River, Off Swann's Point  
 SBE2 South Branch Elizabeth River  
 SBE5 South Branch Elizabeth River, Off VEPCO  
 TF3.3 Rappahannock River, Buoy N40  
 TF4.2 Pamunkey River, Off White House  
 TF5.5 James River, Red Buoy 107  
 WE4.2 Off Mouth of York River

# STATION NAMES, LATITUDES (decimal degrees), LONGITUDES (decimal degrees), TOTAL DEPTH (meters), LATITUDES (degrees, minutes and decimal seconds), and LONGITUDES (degrees, minutes and decimal seconds). These station positions represent target values and are not actual values. They are the values used by the Chesapeake Bay Program as a whole to coordinate data for the stations. Station positions are provided as NAD83 coordinates.

STATION	LATITUDE	LONGITUDE	TOTAL_DEPTH	LATITUDE (DMS)	LONGITUDE (DMS)
CB6.1	37.58833	-76.1625	13.1	37 35' 18"	-77 50' 15"
CB6.4	37.23639	-76.2083	10.5	37 14' 11"	-77 47' 30"
CB7.3E	37.22861	-76.0542	17.8	37 13' 43"	-77 56' 45"

CB7.4	36.99556	-76.0208 13.8	36 59' 44"	-77 58' 45"
LE3.6	37.59667	-76.285 9.8	37 35' 48"	-77 42' 54"
LE5.5	36.99889	-76.3136 21.4	36 59' 48"	-76 18' 12"
LE5.5-W	36.99903	-76.31328 6.0	36 59' 56"	-76 18' 49"
RET3.1	37.92014	-76.8214 5.8	37 55' 12.488"	-77 10' 43.138"
RET4.1	37.52514	-76.8697	37 31' 30.522"	-77 7' 49.131"
RET4.3	37.50681	-76.788 5.2	37 30' 24.522"	-77 12' 43.14"
RET5.2	37.21015	-76.793 8.3	37 12' 36.533"	-77 12' 25.145"
SBE2	36.81265	-76.3058 13.0	36 48' 45.533"	-77 41' 39.212"
SBE5	36.76987	-76.2961 10.0	36 46' 11.534"	-77 42' 14.215"
TF3.3	38.01874	-76.908 6.6	38 1' 7.481"	-77 5' 31.122"
TF4.2	37.57987	-77.0216 6.4	37 34' 47.52"	-78 58' 42.113"
TF5.5	37.31293	-77.2328 9.0	37 18' 46.534"	-78 46' 2.087"
WE4.2	37.24167	-76.3867 14.1	37 14' 30"	-77 36' 48"

Station depths are based on a nine year (1985-1994) averages of Virginia Department of Environmental Quality, water quality hydrographic data collected concurrently with the plankton samples.

#### # METHODOLOGY DESCRIBING CHAIN OF CUSTODY FOR LAB SAMPLES

The phytoplankton field chief is the custodian for all samples collected, and verifies proper labeling of bottles, complete field data entries, the collection of the samples, preservative used and transport to the laboratory. He also supervises the calibration and availability of field equipment. Samples are turned over to the laboratory chief who oversees the sample processing, analysis and recording of the raw data. The principal investigator and laboratory chief for quality assurance routinely check taxon identifications, raw data sheets and other stages of the collection and analysis procedures.

#### # BIOLOGICAL ENUMERATION TECHNIQUES

-Chesapeake Bay Program Sample Analysis Method PH102

Upon return to the laboratory, each 500 ml water sample (fixed with Lugol's solution on station) are preserved with 5 ml of buffered formaldehyde. The 500 ml replicate sample sets are mixed (1000 ml), then 500 ml are withdrawn and allowed to settle undisturbed for 72 hours, the original 500 ml is reduced by careful siphoning to approximately 200-250 ml. The samples are allowed to stand undisturbed for an additional 48 hours and are again siphoned to 20-40 ml concentrates. The final 20-40 ml concentrate is transferred to a previously labeled storage vial, where the label information from the collection bottle has been transferred and verified by the laboratory supervisor. A known volume of the entire concentrate will be placed in an Utermöhl settling chamber for examination with an inverted plankton microscope. If the phytoplankton, and/or silt, density is too great in the final concentrate for clear examination, a known volume of the concentrate is drawn off to provide a sub-sample suitable for analysis. Prior to counting, a work sheet is prepared, where information from the sample vial label is transferred to the raw data sheet and verified. The microscopic examination will be done at 3 magnifications (Marshall and Alden, 1990). At 300X magnification, a combined random field (10) and minimum cell count (200) procedure will be followed where all taxa are counted to the lowest taxonomic category possible. This examination is repeated at 500x magnification for 10 randomly selected fields. Cells not clearly discernable at the 300x magnification are examined at 600X for identification. All species will be counted at only one of these magnifications. In addition, the entire chamber will be scanned at 125X for recording previously unrecorded larger species in the chamber. All phytoplankton categories will be included in this analysis, including colonies and algal filaments at 300x. Calculations will be made from these data at the different magnifications to determine cell concentrations per unit volume (e.g. cells/l). Identification will be based on internationally accepted identification keys, and checked against voucher specimens (e.g. Chesapeake Bay) that are maintained in the ODU phytoplankton analysis laboratory. Samples are archived for a period of one year. Raw data sheets are kept on file.

-Chesapeake Bay Program Analytical Method Code PH102M

Counting Method 102 was slightly modified in January 2005 for better agreement with the new counting methodology adopted in the Maryland Phytoplankton program. Beginning in 2005, at 600X all cells were identified using the following categories:

CENTRIC DIATOMS < 10UM

CRYPTOMONAS <10UM

PENNATE DIATOMS< 10UM

UNIDENTIFIED GREEN CELLS 3-5UM

UNIDENTIFIED MICROPHYTOFLAGELLATES <10UM

All other protocol for sample enumeration remained the same.

-Chesapeake Bay Program analytical Method Code PH103

Beginning in October 2005 the counting enumeration method was slightly modified again to be in agreement with the official Chesapeake Bay program technique for phytoplankton enumeration. The final enumeration protocol for the Virginia program is as follows:

(1) At 300X magnification, a minimum of twenty random fields and 200 cells of taxa > 5 microns in largest dimension will be counted. If 200 cells are not tallied in 10 fields, cells in additional fields will be enumerated until 200 cells have been enumerated. All colonies, trichomes, & filaments are counted at this magnification. Very large (>60 Microns) or rare species (less than 1 cell in less than 10 Grids) not counted in this scan.

(2) At 600X magnification, twenty random fields will be counted for taxa  $\geq 3$  and  $\leq 5$  microns in diameter. No colonies, trichomes or filaments counted. Again all cells were identified using the following categories: CENTRIC DIATOM < 10UM, CRYPTOMONAS <10UM, PENNATE DIATOM < 10UM, UNIDENTIFIED GREEN CELLS 3-5UM UNIDENTIFIED MICROPHYTOFLAGELLATES <10UM

(3) At 125X magnification, the entire chamber will be scanned for taxa which were not enumerated at the other two magnifications.

> Chesapeake Bay Program Sample Analysis Method PP101

Using a Millipore apparatus, a backing 0.45 um nucleopore filter was wetted with distilled water, was placed on the Millipore stem. Then a 0.20 um nucleopore filter, previously stained in an irgalan black solution was placed over the other filter. Two milliliters of the shaken water sample was added to the filter apparatus. Using a pump, and a maximum vacuum of ten centimeter for mercury, the sample was filtered until the meniscus disappears from the top of the filter. The 0.2 um nucleopore filter was removed and placed immediately on a glass slide previously moistened by breath. A drop of immersion oil was placed at the center of the filter, then covered with a cover slip. The slide is examined immediately with a Zeiss Axioskop epifluorescent microscope equipped with a 100 watt mercury lamp and a 100X oil immersion objective. The autotrophic picoplankton were counted using a "green" filter set (g546, FT580, LP590). Count are made on replicate samples and averaged. A minimum of 200 cells and a minimum coverage of 20 field is counted on each slide.

# FORMULAS, CALCULATIONS, AND CONVERSIONS

The following equation was used to convert raw counts to density for each taxon identified for both phytoplankton and picoplankton:

$(\# \text{ cells}/\# \text{ fields counted}) * \text{constant} * (1/\text{concentration} * \text{volume})$

# MONITORING QA/QC PLAN FOR PROJECT

The principal investigator and laboratory chief for quality assurance routinely check taxon identifications, raw data sheets and other stages of the collection and analysis procedures.

# VARIABLE NAMES, MEASUREMENT UNITS AND DESCRIPTIONS

>PARAMETER: COUNT (# of a Phytoplankton Taxon per liter),

-COLLECTION METHODS: At each station, composite phytoplankton samples are taken from above and below the pycnocline. After the pycnocline has been determined at each station, two vertical series of five samples equidistance apart depths are taken between the pycnocline and bottom. Water in each carboy is mixed, then a 500 milliliter sample is taken from each carboy, and is preserved with five milliliters of Lugol's solution. The pre-labeled sample bottles are transported back to the laboratory for analysis.

-SAMPLE PRESERVATION: Lugol's solution

-SAMPLE STORAGE ENVIRONMENT: Room temperature

-TIME IN STORAGE: 1 months

-LABORATORY TECHNIQUE WITH REFERENCES

Marshall, H.G. and R.W. Alden. (1990) A comparison of phytoplankton assemblages and environmental relationships in three estuarine rivers of the lower Chesapeake Bay. *Estuaries*, 13:287-300.

Marshall, H.G. (1994) Chesapeake Bay Phytoplankton: I. Composition. *Proceedings of the Biological Society of Washington*. 107:573-585.

>PARAMETER:COUNT (# Autotrophic Picoplankton per liter)

-COLLECTION METHODS: At each station, composite phytoplankton samples are taken from above and below the pycnocline. After the pycnocline has been determined at each station, two vertical series of five samples equidistance apart depths are taken between the pycnocline and bottom. Water in each carboy is mixed, then a 125 milliliter sample is taken from each carboy, and is preserved with two milliliters of Glutaraldehyde solution. The pre-labeled sample bottles are transported back to the laboratory and placed in a refrigerator for analysis.

-SAMPLE PRESERVATION: Glutaraldehyde solution

-SAMPLE STORAGE ENVIRONMENT: Refrigerator

-TIME IN STORAGE: one week

-LABORATORY TECHNIQUE WITH REFERENCES

Davis, P.G. and J.McN. Sieburth (1982) Differentiation of phototrophic and heterotrophic nanoplankton populations in marine waters by epifluorescence microscopy. *Ann. Inst. Oceanography*. 58:249-260.

Marshall, H.G. (1995) Autotrophic picoplankton distribution and abundance in the Chesapeake Bay, USA *Marine Nature* 4:33-42.

>PARAMETER: LATITUDE and LONGITUDE (in decimal degrees)

COLLECTION METHODS: LORAN-C, NAD27 before July 1995; GPS, NAD83 After July 1995. All positions converted to NAD83 coordinates in 2000.

-SAMPLE PRESERVATIVES: None

-SAMPLE STORAGE ENVIRONMENT: None

-TIME IN STORAGE: None

-LAB TECHNIQUES WITH REFERENCES: Station positions in data set are approximations of actual positions in the field. Station latitudes and longitudes are input into a Loran-C or GPS receiver and sampling begins when boat reaches preprogrammed coordinates. Loran-C is accurate to +/- 1500 ft. The actual Loran or GPS coordinates for each sampling event are not currently recorded in data set.

>PARAMETER: P\_DEPTH

COLLECTION METHODS: Hydrolab CTD

-SAMPLE PRESERVATIVES: None

-SAMPLE STORAGE ENVIRONMENT: None

-TIME IN STORAGE: None

-LAB TECHNIQUES WITH REFERENCES: Water column conductivity is recorded immediately before plankton sampling. P\_DEPTH is set at 0.5 meters above the Pycnocline and is used at the cutoff depth between upper and lower water column composite samples. If a station has no pycnocline the water column is divided in to thirds by total depth and the top third of the water column is treated as the upper water column. The pycnocline is determined as follows:

$((\text{Bottom Conductivity} - \text{Surface Conductivity}) / \text{Bottom Depth}) * 2 = \text{Threshold}$

if Threshold is less than 500, then the station has no pycnocline, else if Threshold is greater than 500, then the pycnocline depth is determined to be the first depth at which the conductivity change is greater than the threshold value.

Units of measurement: Conductivity- uhhos/cm and Depth- meters

>PARAMETER: SALZONE (Salinity zone)

-COLLECTION METHODS: Hydrolab CTD

-SAMPLE PRESERVATIVES: None

-SAMPLE STORAGE ENVIRONMENT: None

-TIME IN STORAGE: None

-LAB TECHNIQUES WITH REFERENCES: Water column salinity, temperature and depth is recorded prior to zooplankton tows. Salinity values are averaged over the layer of the water column and a zone is determined. Salinity Ranges are as follows: Fresh 0-0.5 ppt (F), Oligohaline >0.5-5.0 ppt (O), Mesohaline >5.0-18 ppt (M), and Polyhaline >18-32 ppt (P).

>PARAMETER: TOTAL\_DEPTH

-COLLECTION METHODS: Hydrolab CTD

-SAMPLE PRESERVATIVES: None

-SAMPLE STORAGE ENVIRONMENT: None

-TIME IN STORAGE: None

-LAB TECHNIQUES WITH REFERENCES: Water column salinity, temperature and depth is recorded prior to water collection.

>DATA ENTRY METHOD: Phytoplankton and picoplankton counts are entered using a program written in BASIC. ASCII format files are then converted into SAS datasets and used for statistical analyses. ASCII format data sets for submittal are exported from the SAS datasets.

>DATA VERIFICATION: Double-entry with comparison of two files in SAS. Re-entry until both copies match exactly.

#### # SPECIES INHOUSE CODES AND SCIENTIFIC NAME

The in-house codes used by Old Dominion University are numeric.

>INHOUSE SPECIES LIST: Old Dominion University in-house phytoplankton species codes and Latin Names are as follows:

SPEC_CODE	SOURCE_LBL		
2	Amphiprora sp.	27	Chaetoceros compressus
3	Amphora sp.	28	Chaetoceros concavicornis
4	Amphora proteoides	29	Trachelomonas intermedia
5	Asterionellopsis glacialis	30	Chaetoceros constrictus
6	Asterolampra marylandica	31	Chaetoceros convolutus
7	Asteromphalus sp.	32	Chaetoceros costatus
8	Asteromphalus heptactis	33	Chaetoceros curvisetus
9	Bacteriastrum comosum	34	Chaetoceros danicus
10	Bacteriastrum delicatulum	35	Chaetoceros debilis
11	Bacteriastrum elongatum	36	Chaetoceros decipiens
12	Bacteriastrum hyalinum	37	Chaetoceros densus
13	Odontella sp.	38	Chaetoceros didymus
14	Odontella alternans	39	Chaetoceros didymus var. protuberans
15	Odontella aurita	40	Chaetoceros difficilis
16	Odontella longicuris	41	Chaetoceros gracilis
17	Odontella sinensis	42	Chaetoceros laciniosus
18	Cerataulina pelagica	43	Chaetoceros pelagicus
19	Nitzschia angularis var. affinis	44	Chaetoceros pendulus
20	Chaetoceros sp.	45	Chaetoceros peruvianus
21	Chaetoceros affinis	46	Chaetoceros pseudocurvisetus
22	Chaetoceros affinis var. willei	47	Chaetoceros radians
23	Chaetoceros atlanticus	48	Chaetoceros socialis
24	Chaetoceros borealis	49	Chlorella marina
25	Chaetoceros brevis	50	Chaetoceros tetrastichon
26	Chaetoceros coarctatus	51	Chaetoceros tortissimus
		52	Climacodium sp.

53	<i>Climacodium biconcavum</i>	124	<i>Chroomonas salina</i>
54	<i>Climacodium frauenfeldianum</i>	125	<i>Cryptomonas pseudobaltica</i>
55	<i>Cocconeis</i> sp.	126	<i>Merismopedia elegans</i>
56	<i>Corethron</i> sp.	127	<i>Rhizosolenia hebetata</i>
57	<i>Corethron criophilum</i>	128	<i>Rhizosolenia hebetata</i> f. <i>semispina</i>
58	<i>Coscinodiscus</i> sp.	129	<i>Rhizosolenia imbricata</i>
59	<i>Coscinodiscus centralis</i>	130	<i>Rhizosolenia rhombus</i>
60	<i>Thalassiosira eccentrica</i>	131	<i>Rhizosolenia robusta</i>
61	<i>Coscinodiscus granii</i>	132	<i>Rhizosolenia setigera</i>
62	<i>Thalassiosira leptopus</i>	134	<i>Rhizosolenia styliformis</i>
63	<i>Coscinodiscus marginatus</i>	135	<i>Detonula pumila</i>
64	<i>Coscinodiscus nitidus</i>	136	<i>Skeletonema costatum</i>
65	<i>Coscinodiscus perforatus</i>	137	<i>Stephanopyxis</i> sp.
66	<i>Coscinodiscus radiatus</i>	138	<i>Stephanopyxis palmeriana</i>
68	<i>Chlorella salina</i>	139	<i>Striatella unipunctata</i>
69	<i>Detonula confervacea</i>	140	<i>Surirella</i> sp.
70	<i>Diatoma elongatum</i>	141	<i>Synedra ulna</i>
71	<i>Diatoma hyemale</i>	142	<i>Thalassionema</i> sp.
72	<i>Diploneis</i> sp.	143	<i>Thalassionema nitzschioides</i>
73	<i>Diploneis bombus</i>	144	<i>Thalassiosira decipiens</i>
74	<i>Ditylum brightwellii</i>	145	<i>Thalassiosira delicatula</i>
75	<i>Eucampia cornuta</i>	146	Pennate Diatoms (Unid.) < 20u apical axis
76	<i>Eucampia zodiacus</i>	147	<i>Thalassiosira gravida</i>
77	<i>Fragilaria</i> sp.	148	Pennate Diatoms (Unid.) > 20u apical axis
78	<i>Fragilaria crotonensis</i>	149	<i>Thalassiosira nordenskiöldii</i>
79	<i>Gomphonema</i> sp.	150	<i>Thalassiosira rotula</i>
80	<i>Grammatophora marina</i>	151	<i>Lioloma delicatulum</i>
81	<i>Guinardia flaccida</i>	152	<i>Thalassiothrix frauenfeldii</i>
82	<i>Gyrosigma</i> sp.	153	<i>Thalassiothrix mediterranea</i>
83	<i>Hemiaulus hauckii</i>	154	<i>Tropidoneis</i> sp.
84	<i>Hemiaulus membranaceus</i>	155	Centric Diatoms (Unid.)
85	<i>Hemiaulus sinensis</i>	156	<i>Thalassiosira</i> sp.
86	<i>Pyramimonas amyliifer</i>	157	Centric Diatoms (Unid.) 20u-100u diameter
87	<i>Lauderia annulata</i>	158	Centric Diatoms (Unid.) > 100u diameter
88	<i>Leptocylindrus danicus</i>	159	Dinoflagellates (Unid.)
89	<i>Leptocylindrus minimus</i>	160	<i>Navicula distans</i>
90	<i>Licmophora abbreviata</i>	161	<i>Coscinodiscus rotula</i>
91	<i>Licmophora paradoxa</i>	163	<i>Lauderia</i> sp.
92	<i>Mastogloia</i> sp.	164	<i>Asterionella formosa</i>
93	<i>Melosira</i> sp.	165	<i>Plagiogramma</i> sp.
94	<i>Aulacoseira granulata</i>	166	<i>Aulacoseira distans</i>
95	<i>Melosira hummii</i>	167	<i>Melosira arenaria</i>
96	<i>Paralia sulcata</i>	168	<i>Thalassiosira aestivalis</i>
97	<i>Navicula</i> sp.	169	<i>Dinophysis micropterygia</i>
98	<i>Navicula membranacea</i>	170	<i>Ankistrodesmus convolutus</i>
99	<i>Nitzschia</i> sp.	171	<i>Chaetoceros lorenzianus</i>
100	<i>Cylindrotheca closterium</i>	172	Dinoflagellate cysts (Unid.)
101	<i>Pseudo-nitzschia delicatissima</i>	173	<i>Phalacroma</i> sp.
102	<i>Nitzschia longissima</i>	174	<i>Tetracyclus</i> sp.
103	<i>Pyramimonas grossii</i>	175	<i>Stephanopyxis turris</i>
104	<i>Pseudo-nitzschia pungens</i>	176	<i>Actinopterychus senarius</i>
105	<i>Pseudo-nitzschia seriata</i>	177	<i>Coscinodiscus oculus iridis</i>
106	<i>Plagiogrammopsis vanheurckii</i>	178	<i>Cocconeis clandestina</i>
107	<i>Planktoniella sol</i>	179	<i>Rhizosolenia</i> sp.
108	<i>Pleurosigma</i> sp.	180	<i>Biddulphia biduphiana</i>
109	<i>Pleurosigma angulatum</i>	181	<i>Grammatophora</i> sp.
110	<i>Pleurosigma elongatum</i>	182	<i>Aulacoseira granulata</i> var. <i>angustissima</i>
111	<i>Pleurosigma formosum</i>	183	<i>Oscillatoria</i> sp.
112	<i>Rhizosolenia acuminata</i>	184	<i>Nitzschia frustulum</i>
113	<i>Proboscla alata</i>	185	<i>Odontella rhombus</i> f. <i>trigona</i>
114	<i>Proboscla alata</i> f. <i>curvirostris</i>	186	<i>Diploneis litoralis</i>
115	<i>Proboscla alata</i> f. <i>gracillima</i>	188	<i>Achnanthes subsalsoides</i>
116	<i>Proboscla alata</i> f. <i>indica</i>	189	<i>Glenodinium</i> sp.
117	<i>Rhizosolenia bergonii</i>	190	<i>Grammatophora angulosa</i>
118	<i>Psuedosoleia calcar-avis</i>	191	<i>Coscinodiscus obscurus</i>
119	<i>Rhizosolenia castracanei</i>	192	<i>Tabellaria</i> sp.
120	<i>Guinardia cylindrus</i>	193	<i>Achnanthes</i> sp.
121	<i>Guinardia delicatula</i>	194	<i>Odontella aurita</i> var. <i>obtusa</i>
122	<i>Pyramimonas plurioculata</i>	195	<i>Melosira nummuloides</i>
123	<i>Dactyliosolen fragilissimus</i>	196	<i>Nitzschia angularis</i>

197	<i>Thalassiosira hyalina</i>	269	<i>Coscinodiscus gigas</i> var. <i>praetexta</i>
198	<i>Achnanthes taeniata</i>	270	<i>Mastogloia rostrata</i>
199	<i>Bacteriastrum</i> sp.	271	<i>Pleurosigma directum</i>
200	<i>Protoperidinium hirobis</i>	272	<i>Amphiprora alata</i>
201	<i>Gymnodinium dissimile</i>	273	<i>Hemidiscus cuneiformis</i>
202	<i>Gyrodinium fusiforme</i>	274	<i>Tropidoneis lepidoptera</i>
203	<i>Prorocentrum obtusum</i>	275	<i>Biddulphia tridentata</i>
204	<i>Licmophora</i> sp.	276	<i>Triceratium reticulatum</i>
205	<i>Amphidinium lacustre</i>	277	<i>Campylodiscus limbatus</i>
206	<i>Melosira moniliformis</i>	278	<i>Cyclotella meneghiniana</i>
207	<i>Licmophora gracilis</i>	279	<i>Thalassiosira pseudonana</i>
208	<i>Diploneis subcincta</i>	280	<i>Coscinodiscus gigas</i>
209	<i>Chaetoceros teres</i>	281	<i>Coscinodiscus asteromphalus</i>
210	<i>Nitzschia vitrea</i>	283	<i>Chaetoceros simplex</i>
211	<i>Aulacoseira islandica</i>	284	<i>Chaetoceros diadema</i>
212	<i>Dinophysis hastata</i>	285	<i>Cymatosira belgica</i>
213	<i>Pleurosigma naviculaceum</i>	286	<i>Dactyliosolen antarcticus</i>
214	<i>Achnanthes delicatula</i>	287	<i>Diploneis crabro</i>
215	<i>Prorocentrum triestinum</i>	288	<i>Diploneis crabro</i> var. <i>pandura</i>
216	<i>Anthosphaera robusta</i>	289	<i>Eunotia</i> sp.
217	<i>Synedra</i> sp.	290	<i>Eunotia bidentula</i>
218	<i>Cocconeis costata</i>	291	<i>Rhabdosphaera hispida</i>
219	<i>Cocconeis distans</i>	292	<i>Podosira stelligera</i>
220	<i>Gephyrocapsa caribbeanica</i>	293	<i>Rhizosolenia temperei</i>
221	<i>Chaetoceros messanensis</i>	294	<i>Striatella interrupta</i>
222	<i>Gephyrocapsa ericsonii</i>	295	<i>Synedra gaillonii</i>
223	<i>Diatoma</i> sp.	296	<i>Synedra fulgens</i>
224	<i>Toxarium undulatum</i>	297	<i>Crucigenia tetrapedia</i>
225	<i>Ophiaster hydroideus</i>	298	<i>Synedra toxoneides</i>
226	<i>Melosira islandica</i> f. <i>curvata</i>	299	<i>Synedra crystallina</i>
228	<i>Acantheoca aculeata</i>	300	<i>Amphidinium</i> sp.
229	<i>Odontella mobiliensis</i>	301	<i>Ceratium</i> sp.
230	<i>Achnanthes lemmermannii</i>	302	<i>Ceratium arcticum</i>
231	<i>Florisphaera profunda</i>	303	<i>Ceratium azoricum</i>
232	<i>Fragilariopsis oceanica</i>	304	<i>Ceratium candelabrum</i>
233	<i>Chaetoceros tenuissimus</i>	305	<i>Ceratium extensum</i>
234	<i>Thalassiosira baltica</i>	306	<i>Ceratium furca</i>
235	<i>Thalassiosira bioculata</i>	307	<i>Ceratium fuscum</i>
236	<i>Syracosphaera corolla</i>	308	<i>Ceratium lineatum</i>
237	<i>Syracosphaera histrica</i>	309	<i>Ceratium longipes</i>
238	<i>Odontella granulata</i>	310	<i>Ceratium macroceros</i>
239	<i>Protoperidinium sub-curvipes</i>	311	<i>Ceratium massiliense</i>
240	<i>Stephanopyxis nipponica</i>	312	<i>Ceratium minutum</i>
241	<i>Protoperidinium mite</i>	313	<i>Ceratium pentagonum</i>
242	<i>Grammatophora serpentina</i>	314	<i>Pediastrum duplex</i> var. <i>reticulatum</i>
243	<i>Prorocentrum rotundatum</i>	315	<i>Ceratium tripos</i>
244	<i>Prorocentrum ovum</i>	316	<i>Ornithocercus</i> sp.
245	<i>Azpeitia tabularis</i>	317	<i>Nostoc</i> sp.
247	<i>Fragilaria construens</i>	318	<i>Chaetoceros neapolitanus</i>
248	<i>Chaetoceros subtilis</i>	319	<i>Skeletonema potamos</i>
249	<i>Achnanthes longipes</i>	320	<i>Dinophysis</i> sp.
250	<i>Prorocentrum cordatum</i>	321	<i>Dinophysis acuminata</i>
251	<i>Chaetoceros wighamii</i>	322	<i>Dinophysis acuta</i>
252	<i>Thalassiothrix longissima</i>	323	<i>Dinophysis arctica</i>
253	<i>Rhaphoneis</i> sp.	324	<i>Dinophysis caudata</i>
254	<i>Amphidinium operculatum</i>	325	<i>Dinophysis norvegica</i>
255	<i>Leptocylindrus mediterraneus</i>	326	<i>Dinophysis ovum</i>
256	<i>Fragilaria pinnata</i>	327	<i>Dinophysis punctata</i>
257	<i>Chaetoceros similis</i>	328	<i>Dinophysis schuettii</i>
258	<i>Bleakeleya notata</i>	329	<i>Agmenellum thermale</i>
259	<i>Thalassiosira antarctica</i>	330	<i>Prorocentrum compressum</i>
260	<i>Membraneis challengerii</i>	331	<i>Eutreptia</i> sp.
261	<i>Cyclotella striata</i>	332	<i>Gonyaulax digitale</i>
262	<i>Aulacoseira italica</i>	333	<i>Gonyaulax polygramma</i>
263	<i>Coscosira polychorda</i>	334	<i>Gonyaulax spinifera</i>
264	<i>Melosira jurgensii</i>	335	<i>Gonyaulax triacantha</i>
265	<i>Caloneis fusioides</i>	336	<i>Gonyaulax</i> sp.
266	<i>Asteromphalus roperianus</i>	337	<i>Gymnodinium</i> sp.
267	<i>Prorocentrum nanum</i>	338	<i>Gymnodinium costatum</i>
268	<i>Licmophora flabellata</i>	339	<i>Gymnodinium rhomboides</i>



340	<i>Gymnodinium simplex</i>	411	<i>Gymnodinium splendens</i>
341	<i>Gyrodinium sp.</i>	412	<i>Katodinium rotundatum</i>
342	<i>Gyrodinium lacryma</i>	413	<i>Pavillardinium spinosa</i>
343	<i>Noctiluca scintillans</i>	414	<i>Oxytoxum obliquum</i>
344	<i>Fragilaria striatula</i>	415	<i>Ceratium setaceum</i>
345	<i>Pediastrum biradiatum</i>	416	<i>Ceratium teres</i>
346	<i>Oxytoxum gladiolus</i>	417	<i>Protoperidinium fimbriatum</i>
347	<i>Oxytoxum milneri</i>	418	<i>Amphidinium turbo</i>
348	<i>Oxytoxum reticulatum</i>	419	<i>Ceratium belone</i>
349	<i>Oxytoxum scolopax</i>	420	<i>Diploneis smithii</i>
350	<i>Oxytoxum sphaeroideum</i>	421	<i>Oxytoxum longiceps</i>
351	<i>Oxytoxum variabile</i>	422	<i>Amphidinium steinii</i>
352	<i>Dissodium asymmetricum</i>	423	<i>Prorocentrum aporum</i>
353	<i>Pyrophacus sp.</i>	424	<i>Gymnodinium marinum</i>
354	<i>Oblea rotunda</i>	425	<i>Pavillardinium sp.</i>
355	<i>Protoperidinium sp.</i>	426	<i>Calciosolenia granii</i>
356	<i>Protoperidinium breve</i>	427	<i>Pyrocystis fusiformis</i>
357	<i>Protoperidinium cerasus</i>	428	Green cells (
358	<i>Protoperidinium conicoides</i>	429	Green cells (3-5 microns)
359	<i>Protoperidinium depressum</i>	430	Green cells (5-10 microns)
360	<i>Protoperidinium leonis</i>	431	<i>Chlorella sp.</i>
361	<i>Protoperidinium oceanicum</i>	432	<i>Oxytoxum belgicae</i>
362	<i>Protoperidinium ovatum</i>	433	<i>Amphidinium sphenoides</i>
363	<i>Protoperidinium pallidum</i>	434	<i>Mallomonas sp.</i>
364	<i>Protoperidinium pellucidum</i>	435	<i>Calycomonas gracilis</i>
365	<i>Protoperidinium pentagonum</i>	436	<i>Oxytoxum crassum</i>
366	<i>Protoperidinium punctulatum</i>	437	<i>Protoperidinium claudicans</i>
367	<i>Thalassiosira subtilis</i>	438	<i>Dinophysis parvula</i>
368	<i>Crucigenia irregularis</i>	439	<i>Coscinodiscus nobilis</i>
369	<i>Podolampas bipes</i>	440	<i>Amphidoma acuminata</i>
370	<i>Podolampas elegans</i>	441	<i>Coscinodiscus subbulliens</i>
371	<i>Podolampas palmipes</i>	442	<i>Prorocentrum lima</i>
372	<i>Prorocentrum sp.</i>	443	<i>Ceratium horridum</i>
373	<i>Prorocentrum dentatum</i>	444	<i>Glenodinium gymnodinium</i>
374	<i>Prorocentrum micans</i>	445	<i>Oxytoxum laticeps</i>
375	<i>Prorocentrum minimum</i>	446	<i>Actinoptychus splendens</i>
376	<i>Prorocentrum rostratum</i>	447	<i>Heterocapsa triquetra</i>
377	<i>Prorocentrum scutellum</i>	448	<i>Ceratium longinum</i>
378	<i>Nitzschia gracillima</i>	449	<i>Ceratium trichoceros</i>
379	<i>Coscinodiscus sublineatus</i>	450	<i>Odontella regia</i>
380	<i>Anacystis sp.</i>	451	<i>Gonyaulax conjuncta</i>
381	<i>Gymnodinium arcticum</i>	452	<i>Glenodinium danicum</i>
382	<i>Ceratium tripos var. atlanticum</i>	453	<i>Gonyaulax longispina</i>
383	<i>Dinophysis rotundata</i>	454	<i>Rhabdosphaera longistylis</i>
384	<i>Dinophysis fortii</i>	455	<i>Navicula paleralis</i>
385	<i>Oxytoxum sceptrum</i>	456	<i>Protoperidinium bipes</i>
386	<i>Cladopyxis brachiolata</i>	457	<i>Amphora crassa</i>
387	<i>Prorocentrum vaginulum</i>	458	<i>Cochlodinium sp.</i>
388	<i>Gonyaulax polyedra</i>	459	<i>Oxytoxum gracile</i>
389	<i>Ceratium arietinum</i>	460	<i>Heteromastix pyriformis</i>
390	<i>Amphidinium schroederi</i>	461	<i>Protoperidinium pyriforme</i>
391	<i>Amphidinium bipes</i>	462	<i>Scrippsiella trochoidea</i>
393	<i>Protoperidinium conicum</i>	463	<i>Dinophysis lachmannii</i>
394	<i>Triceratium acutum</i>	464	<i>Cystodinium sp.</i>
395	<i>Podolampas sp.</i>	465	<i>Pyramimonas obovata</i>
396	<i>Amphidinium globosum</i>	466	<i>Gymnodinium conicum</i>
397	<i>Gonyaulax minima</i>	467	<i>Amphidinium ovoideum</i>
398	<i>Oxytoxum mitra</i>	468	<i>Gymnodinium punctatum</i>
399	<i>Oxytoxum diploconus</i>	469	<i>Amphidoma sp.</i>
400	<i>Ceratium longirostrum</i>	470	<i>Oxytoxum globosum</i>
401	<i>Gonyaulax birostris</i>	471	<i>Ceratocorys horrida</i>
402	<i>Monodus sp.</i>	472	<i>Podolampas spinifera</i>
403	<i>Oxytoxum constrictum</i>	473	<i>Prorocentrum gracile</i>
404	<i>Dinophysis minuta</i>	474	<i>Protoperidinium globulum</i>
405	<i>Protoperidinium brevipes</i>	475	<i>Amphidinium acutum</i>
406	<i>Prorocentrum balticum</i>	476	<i>Gonyaulax diacantha</i>
407	<i>Amphidinium latum</i>	477	<i>Pyrocystis lunula</i>
408	<i>Bellerophon horologicalis</i>	478	<i>Protoperidinium granii</i>
409	<i>Gonyaulax minuta</i>	479	<i>Amphidinium glaucum</i>
410	<i>Heteraulacus polyedricus</i>	480	<i>Amphidinium wislouchi</i>

481	<i>Protopteridinium diabolum</i>	551	<i>Syracosphaera</i> sp.
482	<i>Gonyaulax unicornis</i>	552	<i>Rhabdosphaera</i> sp.
483	<i>Cochlodinium constrictum</i>	553	<i>Cyclotella atomus</i>
484	<i>Diplopsalis lenticula</i>	554	<i>Amphora egregia</i> var. <i>interrupta</i>
485	<i>Gyrodinium spirale</i>	555	<i>Pontosphaera</i> sp.
486	<i>Protopteridinium nudum</i>	556	<i>Discosphaera</i> sp.
487	<i>Gyrodinium undulans</i>	557	<i>Ceratium hirundinella</i>
488	<i>Amphidinium crassum</i>	558	<i>Surirella cruciata</i>
489	<i>Protopteridinium orbiculare</i>	559	<i>Hantzschia marina</i>
490	<i>Protopteridinium steinii</i>	560	<i>Calycomonas wulffii</i>
491	<i>Protopteridinium subinerme</i>	561	<i>Chrysoococcus minutus</i>
492	<i>Dinophysis sphaerica</i>	562	<i>Tribonema affine</i>
493	<i>Protopteridinium excentricum</i>	563	<i>Tribonema monochloron</i>
494	<i>Cladopyxis setifera</i>	564	<i>Heteromastix rotunda</i>
495	<i>Dinophysis exigua</i>	565	<i>Pavlova salina</i>
496	<i>Protopteridinium minutum</i>	566	<i>Tetraselmis maculata</i>
497	<i>Protopteridinium oblongum</i>	567	<i>Pyramimonas micron</i>
498	<i>Ceratium breve</i>	568	<i>Phacus curvicauda</i>
499	<i>Protopteridinium abei</i>	569	<i>Phacus</i> sp.
500	<i>Acanthoica</i> sp.	570	<i>Phacus longicauda</i>
501	<i>Acanthoica acanthifera</i>	571	<i>Phacus lemmermanni</i>
502	<i>Acanthoica acanthos</i>	572	<i>Euglena proxima</i>
503	<i>Gonyaulax excavata</i>	573	<i>Euglena mutabilis</i> var. <i>mainxi</i>
504	<i>Acanthoica quattrosolina</i>	574	<i>Euglena pumila</i>
505	<i>Anoplosolenia brasiliensis</i>	575	<i>Ceratium declinatum</i>
506	<i>Chilomonas marina</i>	576	<i>Protopteridinium crassipes</i>
507	<i>Anthosphaera</i> sp.	577	<i>Auliscus caelatus</i>
508	<i>Anthosphaera quadriconu</i>	578	<i>Ochromonas caroliniana</i>
509	<i>Chroomonas vectensis</i>	579	<i>Gyrodinium dominans</i>
510	<i>Cryptomonas stigmatica</i>	580	<i>Tetraselmis gracilis</i>
511	<i>Calciosolenia murrayi</i>	581	<i>Histoneis variabilis</i>
512	<i>Calyptrosphaera</i> sp.	582	<i>Ornithocercus quadratus</i>
513	<i>Procentrum cassubicum</i>	583	<i>Ornithocercus steinii</i>
514	<i>Calyptrosphaera oblonga</i>	584	<i>Ornithocercus thumii</i>
515	<i>Emiliana huxleyi</i>	585	<i>Oxytoxum coronatum</i>
516	<i>Coccolithus pelagicus</i>	586	<i>Oxytoxum curvatum</i>
517	<i>Hymenomonas carterae</i>	587	<i>Auliscus sculptus</i>
518	<i>Cyclococcolithus fragilis</i>	588	<i>Ceratium ranipes</i>
519	<i>Cyclococcolithus leptoporus</i>	589	<i>Protopteridinium nipponicum</i>
520	<i>Discosphaera tubifer</i>	590	<i>Protopteridinium achromaticum</i>
521	<i>Gephyrocapsa oceanica</i>	591	<i>Dinophysis tripos</i>
522	<i>Halopappus adriaticus</i>	592	<i>Polykrikos kofoidii</i>
523	<i>Pseudo-nitzschia subpacific</i>	593	<i>Procentrum maximum</i>
524	<i>Calciosolenia</i> sp.	594	<i>Podolampas curvatus</i>
525	<i>Pyramimonas</i> sp.	595	<i>Pyrocystis fusiformis</i> f. <i>biconica</i>
526	<i>Spirulina</i> sp.	596	<i>Pyrocystis robusta</i>
527	<i>Michaelsarsia elegans</i>	597	<i>Pyrocystis pseudonociluca</i>
528	<i>Desmidium</i> sp.	598	<i>Katodinium asymmetricum</i>
529	<i>Dinophysis monacantha</i>	599	<i>Bacteriastrum hyalinum</i> var. <i>princeps</i>
530	<i>Pontosphaera syracusana</i>	600	<i>Tetraselmis</i> sp.
531	<i>Rhabdosphaera clavigera</i>	601	<i>Triceratium favus</i>
532	<i>Chaetoceros rostratus</i>	602	<i>Tropidoneis seriata</i>
533	<i>Rhabdosphaera styliifer</i>	603	<i>Chrysochromulina</i> sp.
534	<i>Crucigenia</i> sp.	604	<i>Actinoptychus vulgaris</i>
535	<i>Scyphosphaera apsteinii</i>	605	<i>Amphora acuta</i>
536	<i>Umbellosphaera tenuis</i>	606	<i>Amphora binodis</i>
537	<i>Staurastrum quadricuspidatum</i>	607	<i>Amphora grevilleana</i> var. <i>contracta</i>
538	<i>Pyramimonas torta</i>	608	<i>Amphora coffeaeformis</i>
539	<i>Triceratium formosum pentagonal</i>	609	<i>Amphora cuneata</i>
540	<i>Syracosphaera mediterranea</i>	610	<i>Amphora terroris</i>
541	<i>Syracosphaera molischii</i>	611	<i>Amphora obtusa</i>
542	<i>Syracosphaera pirus</i>	612	<i>Amphora ostrearia</i>
543	<i>Syracosphaera pulchra</i>	613	<i>Amphora ovalis</i>
544	<i>Schizothrix tenerima</i>	614	<i>Amphora peragalli</i>
545	<i>Navicula directa</i>	615	<i>Amphora proteus</i>
546	<i>Umbellosphaera irregularis</i>	616	<i>Amphora robusta</i>
547	<i>Umbilicosphaera hulbertiana</i>	617	<i>Gyrosigma wansbeckii</i>
548	<i>Coccolithophores (Unid.)</i>	618	<i>Asterionellopsis kariana</i>
549	<i>Katodinium</i> sp.	619	<i>Asteromphalus flabellatus</i>
550	<i>Thalassiosira nana</i>	620	<i>Bacillaria paxillifer</i>

621	<i>Bacteriastrum furcatum</i> Shadbolt	693	<i>Amphiprora gigantea</i> var. <i>sulcata</i>
622	<i>Caloneis staurophora</i>	694	<i>Licmophora paradoxa</i> var. <i>tincta</i>
623	<i>Caloneis wardii</i>	695	<i>Navicula cuspidata</i> var. <i>ambigua</i>
624	<i>Mastogloia braunii</i>	696	<i>Synedra superba</i>
625	<i>Ochromonas</i> sp.	698	<i>Amphora laevis</i>
626	<i>Chaetoceros diversus</i>	699	<i>Cocconeis scutellum</i>
627	<i>Cocconeis molesta</i> var. <i>crucifera</i>	700	<i>Scenedesmus armatus</i>
628	<i>Cocconeis pinnata</i>	701	<i>Dictyocha fibula</i>
629	<i>Coscinodiscus apiculiferus</i>	702	<i>Scenedesmus dimorphus</i>
630	<i>Coscinodiscus argus</i>	703	<i>Scenedesmus abundans</i>
631	<i>Coscinodiscus granulatus</i>	704	<i>Dactylococcopsis raphidioides</i>
632	<i>Stellarima microtrias</i>	705	<i>Dinophysis diegensis</i>
633	<i>Fragilariopsis cylindrus</i>	706	<i>Distephanus speculum</i>
634	<i>Cymatosira lorenziana</i>	707	<i>Asterionella</i> sp.
635	<i>Glyphodesmis distans</i>	708	<i>Mesocena polymorpha</i>
636	<i>Gyrosigma hippocampus</i>	709	<i>Silicoflagellates</i> (Unid.)
637	<i>Mastogloia smithii</i>	710	<i>Chlorophyceans</i> (Unid.)
638	<i>Melosira dubia</i>	711	<i>Crucigenia quadrata</i>
639	<i>Navicula abrupta</i>	712	<i>Ceratium limulus</i>
640	<i>Navicula annulata</i>	713	<i>Skeletonema</i> sp.
641	<i>Navicula cancellata</i>	714	<i>Helicotheca tamesis</i>
642	<i>Navicula clavata</i>	715	<i>Gomphosphaeria</i> sp.
643	<i>Navicula forcipata</i>	716	<i>Melosira islandica</i> var. <i>helvetica</i>
644	<i>Navicula inserata</i>	717	<i>Podosira</i> sp.
645	<i>Navicula lundstroemii</i>	718	<i>Ebria tripartita</i>
646	<i>Navicula opima</i>	719	<i>Stauroneis salina</i>
647	<i>Navicula palpebralis</i>	720	<i>Synura</i> sp.
648	<i>Navicula praetexta</i>	721	<i>Cerataulus radiatus</i>
649	<i>Navicula pusilla</i>	722	<i>Asterionella notata</i>
650	<i>Navicula transitans</i> var. <i>asymmetrica</i>	723	<i>Stauroneis</i> sp.
651	<i>Nitzschia constricta</i>	724	<i>Syracosphaera prolongata</i>
652	<i>Nitzschia distans</i>	725	<i>Trachelomonas acanthostoma</i>
653	<i>Nitzschia lorenziana</i> var. <i>densistriata</i>	726	<i>Trachelomonas charkowiensis</i>
654	<i>Nitzschia insignis</i>	727	<i>Trachelomonas hispida</i>
655	<i>Nitzschia lorenziana</i> var. <i>incerta</i>	728	<i>Trachelomonas volvocina</i> var. <i>punctata</i>
657	<i>Nitzschia panduriformis</i>	729	<i>Trachelomonas</i> sp.
658	<i>Nitzschia sigma</i>	730	<i>Protoperidinium avellana</i>
659	<i>Nitzschia sigma</i> var. <i>intercedens</i>	731	<i>Protoperidinium quarnerense</i>
660	<i>Nitzschia sigma</i> var. <i>rigida</i>	732	<i>Protoperidinium roseum</i>
661	<i>Nitzschia socialis</i>	733	<i>Dinophysis schroederi</i>
662	<i>Meridion circulare</i>	734	<i>Dinophysis sacculus</i>
663	<i>Pinnularia trevelyana</i>	735	<i>Cladopyxis</i> sp.
664	<i>Pinnularia rectangulata</i>	736	<i>Protoperidinium thorianum</i>
665	<i>Plagiogramma staurophorum</i>	737	<i>Gymnodinium variabile</i>
666	<i>Pleurosigma aestuarii</i>	738	<i>Thalassiothrix</i> sp.
667	<i>Pleurosigma hamuliferum</i>	739	<i>Gonyaulax fragilis</i>
668	<i>Pleurosigma nicobaricum</i>	740	<i>Actinastrum</i> sp.
669	<i>Pleurosigma normanii</i>	741	<i>Oxytoxum mediterraneum</i>
670	<i>Pleurosigma obscurum</i>	742	<i>Amphidinium carterae</i>
671	<i>Pleurosigma angulatum</i> var. <i>strigosa</i>	743	<i>Ceratium buceros</i> var. <i>tenuis</i>
672	<i>Hemiaulus indicus</i>	744	<i>Oxytoxum parvum</i>
673	<i>Rhaphoneis amphicerus</i>	745	<i>Dinophysis dentata</i>
674	<i>Delphineis surirella</i>	746	<i>Ceratium contortum karstenii</i>
675	<i>Rhizosolenia formosa</i>	747	<i>Dinophysis pulchella</i>
676	<i>Amphora marina</i>	748	<i>Hemiaulus</i> sp.
677	<i>Nitzschia clausii</i>	749	<i>Protoperidinium divergens</i>
678	<i>Scoliotropis latestriata</i>	750	<i>Amphisolenia</i> sp.
679	<i>Stauroneis amphioxys</i>	751	<i>Ankistrodesmus</i> sp.
680	<i>Stenopteroberia anceps</i>	752	<i>Pyrophacus horologium</i>
681	<i>Surirella pandura contracta</i>	753	<i>Ceratium gallicum</i>
682	<i>Closterium</i> sp.	754	<i>Oxytoxum turbo</i>
683	<i>Synedra robusta</i>	755	<i>Amphisolenia bidentata</i>
684	<i>Amphora costata</i>	756	<i>Pavillardinium biconica</i>
686	<i>Trinacria regina</i>	757	<i>Cladopyxis caryophyllum</i>
687	<i>Diploneis constricta</i>	758	<i>Dinophysis baltica</i>
688	<i>Mastogloia cocconeiformis</i>	759	<i>Gonyaulax monocantha</i>
689	<i>Crucigenia apiculata</i>	760	<i>Triceratium</i> sp.
690	<i>Coscinodiscus wailesii</i>	761	<i>Oxytoxum elegans</i>
691	<i>Achnanthes fimbriata</i>	762	<i>Ceratium schroeteri</i>
692	<i>Dimerogramma</i> sp.	763	<i>Chroococcus limneticus</i>

764	<i>Amphidinium acutissimum</i>	835	<i>Diplopetopsis minor</i>
765	<i>Gymnodinium pygmaeum</i>	836	<i>Thalassiosira oestrupii</i> var. <i>venrickae</i>
766	<i>Nitzschia spathulata</i>	837	<i>Cyclotella glomerata</i>
767	<i>Striatella</i> sp.	838	<i>Calycomonas</i> sp.
768	<i>Gymnodinium grammaticum</i>	839	Chrysophyceans (Unid.)
769	<i>Ceratium buceros</i>	840	<i>Cyclotella caspia</i>
770	<i>Gymnodinium flavum</i>	841	<i>Oscillatoria trichomes</i>
771	<i>Ceratium pulchellum</i> f. <i>semipulchellum</i>	842	<i>Amphidinium extensum</i>
772	<i>Amphisolenia globifera</i>	843	<i>Tribonema</i> sp.
773	<i>Actinastrum hantzschii</i>	844	<i>Chaetoceros neogracilis</i>
774	<i>Gymnodinium coeruleum</i>	845	<i>Rhopalodia gibberula</i>
775	<i>Gymnodinium danicans</i>	846	<i>Cryptomonas</i> sp. 2
776	<i>Gyrodinium estuariale</i>	847	<i>Rhopalodia</i> sp.
777	<i>Oxytoxum</i> sp.	848	<i>Gymnodinium</i> sp. b
778	<i>Protoperidinium biconicum</i>	849	<i>Gyrosigma spenceri</i>
779	<i>Protoperidinium brochii</i>	850	<i>Euglena</i> sp.
780	<i>Protoperidinium grande</i>	851	<i>Eutreptia marina</i>
781	<i>Gymnodinium nelsonii</i>	852	<i>Eutreptia viridis</i>
782	<i>Protoperidinium pendunculatum</i>	853	<i>Calothrix</i> sp.
783	<i>Protoperidinium solidicorne</i>	854	<i>Eutreptia lanowii</i>
784	<i>Protoperidinium sphaericum</i>	855	<i>Euglena acus</i>
785	<i>Amphidinium klebsii</i>	856	<i>Euglena agilis</i>
786	<i>Amphidinium lanceolatum</i>	857	<i>Euglena ehrenbergii</i>
787	<i>Ceratium carriense</i>	858	<i>Euglena deses</i>
788	<i>Ceratium contortum</i>	859	<i>Euglena fusca</i>
789	<i>Ceratium contrarium</i>	860	<i>Chilomonas</i> sp.
790	<i>Ceratium digitatum</i>	861	<i>Chroomonas</i> sp.
791	<i>Ceratium geniculatum</i>	862	<i>Cryptomonas</i> sp.
792	<i>Ceratium kofoidii</i>	863	<i>Olisthodiscus luteus</i>
793	<i>Ceratium pavillardii</i>	864	<i>Synedra acus</i>
794	<i>Gonyaulax apiculata</i>	865	<i>Chroomonas amphioxeia</i>
795	<i>Gonyaulax diegensis</i>	866	<i>Cryptomonas rostellata</i>
796	<i>Gonyaulax monilata</i>	867	<i>Cladopyxis claytonii</i>
797	<i>Gymnodinium breve</i>	868	<i>Oocystis</i> sp.
798	<i>Protoperidinium globulus</i> var. <i>ovatum</i>	869	<i>Asterionella gracillima</i>
799	<i>Gymnodinium situla</i>	870	<i>Oscillatoria limnetica</i>
800	<i>Oscillatoria erythraea</i>	871	<i>Pediastrum</i> sp.
801	<i>Lithodesmium</i> sp.	872	<i>Scenedesmus</i> sp.
802	<i>Histoneis longicollis</i>	873	<i>Scenedesmus quadricauda</i>
803	Choanoflagellates (Unid.)	874	<i>Staurastrum manfeldtii</i> var. <i>flumenense</i>
804	Micro-phytoflagellates (Unid.) < 10 $\mu$	875	<i>Staurastrum leptocladum</i> var. <i>insigne</i>
805	Micro-phytoflagellates (Unid.) > 10 $\mu$	876	<i>Scenedesmus acuminatus</i>
806	<i>Richelia intracellularis</i>	877	<i>Staurastrum</i> sp.
807	<i>Coscinodiscus concinnus</i>	878	<i>Crucigenia fenestrata</i>
808	<i>Amphidinium longum</i>	879	<i>Kirchneriella</i> sp.
809	<i>Anabaena</i> sp.	880	<i>Tetraedron minimum</i>
810	<i>Spirulina subsala</i>	881	<i>Crucigenia crucifera</i>
811	<i>Pyrocystis</i> sp.	882	<i>Pediastrum duplex</i>
812	<i>Entophysalis deusta</i>	883	<i>Tetraedron trigonum</i> var. <i>gracile</i>
813	<i>Anacystis cyanea</i>	884	<i>Kirchneriella lunaris</i>
814	<i>Chroococcus turgidus</i>	885	<i>Tetrastrum staurogeniaeforme</i>
815	Blue Green Single Spheres (Unid.)	886	<i>Arthrodesmus</i> sp.
816	Blue Green Trichomes (Unid.)	887	<i>Chlorella vulgaris</i>
817	<i>Agmenellum quadruplicatum</i>	888	<i>Chlorella saccharophila</i> var. <i>ellipsoidea</i>
818	<i>Gomphosphaeria aponina</i>	889	<i>Nannochloris atomus</i>
819	<i>Johannesbaptistia pellucida</i>	890	<i>Enteromorpha intestinalis</i>
820	<i>Nostoc commune</i>	891	<i>Monodus guttula</i>
821	<i>Microcystis aeruginosa</i>	892	<i>Oedogonium</i> sp.
822	<i>Synechococcus</i> sp.	893	<i>Tetraedron</i> sp.
823	<i>Oscillatoria submembranacea</i>	894	<i>Tetraedron muticum</i>
824	<i>Schizothrix calcicola</i>	895	<i>Chrysochromulina minor</i>
825	<i>Nodularia harveyana</i>	896	<i>Ochromonas variabilis</i>
826	<i>Oscillatoria lutea</i>	897	<i>Ochromonas minuscula</i>
827	<i>Calothrix parietina</i>	898	<i>Chromulina parvula</i>
828	<i>Microcystis</i> sp.	899	<i>Calycomonas ovalis</i>
829	<i>Microcoleus lyngbyaceus</i>	900	<i>Pediastrum simplex</i>
830	<i>Schizothrix arenaria</i>	901	<i>Cyclotella stylum</i>
831	<i>Chroococcus</i> sp.	902	<i>Diploneis obliqua</i>
833	<i>Biddulphia reticulata</i>	903	<i>Diploneis suborbicularis</i>
834	Epiphytic diatoms	904	<i>Gyrosigma balticum silimis</i>

905	<i>Mastogloia gibbosa</i>	977	<i>Chaetoceros muelleri</i>
906	<i>Mastogloia pumila</i>	978	<i>Dinophysis doryphorum</i>
907	<i>Tabellaria fenestrata</i>	979	<i>Oxytoxum tessellatum</i>
908	<i>Navicula arenaria</i>	980	<i>Tribonema minus</i>
909	<i>Rhabdonema arcuatum</i>	981	<i>Lemma minor</i>
910	<i>Gyrosigma spenceri</i> var. <i>nodiferum</i>	982	<i>Campsopogan coeruleus</i>
911	<i>Gyrosigma macrum</i>	983	<i>Merismopedia</i> sp.
912	<i>Dimerogramma minor</i>	984	<i>Pyrocystis hamulus</i>
914	<i>Pronoclituca</i> sp.	985	<i>Ornithocercus magnificus</i>
915	<i>Gyrosigma fasciola</i>	986	<i>Arthrodesmus sublatus</i>
916	<i>Pinnularia</i> sp.	987	<i>Scenedesmus bijuga</i>
917	<i>Surirella ovalis</i>	988	<i>Ankistrodesmus falcatus</i>
918	<i>Amphora arenaria</i>	989	<i>Ankistrodesmus falcatus</i> var. <i>acicularis</i>
919	<i>Kirchneriella contorta</i>	990	<i>Tetraedron trigonum</i> var. <i>setigerum</i>
920	<i>Cocconeis scutellum</i> var. <i>ornata</i>	991	<i>Chlamydomonas</i> sp.
921	<i>Merismopedia punctata</i>	992	<i>Coscinodiscus cinctus</i>
922	<i>Navicula maculata</i>	993	<i>Aulacodiscus</i> sp.
923	<i>Nitzschia bilobata</i>	994	<i>Pseudotetraedron neglectum</i>
924	<i>Nitzschia obtusa</i> var. <i>scalpelliformis</i>	995	<i>Porosira gracialis</i>
925	<i>Pleurosigma strigosum</i>	996	<i>Tetraedron trigonum</i>
926	<i>Staurastrum paradoxum</i>	997	<i>Schroederia setigera</i>
927	<i>Amphora gigantea</i>	998	<i>Scenedesmus bijuga</i> var. <i>alternans</i>
928	<i>Amphora szaboi</i>	999	<i>Staurastrum americanum</i>
929	<i>Caloneis subsalina</i>	1000	<i>Staurastrum leptocladum</i>
930	<i>Campylosira cymbelliformis</i>	1001	<i>Micractinium</i> sp.
931	<i>Fragilaria capucina</i>	1002	<i>Synedra closterioides</i>
932	<i>Fragilaria hyalina</i>	1003	<i>Cymbella tumida</i>
933	<i>Navicula humerosa</i>	1004	<i>Dinobryon</i> sp.
934	<i>Merismopedia tenuissima</i>	1005	<i>Gomphonema sphaerophorum</i>
935	<i>Anabaena spiroides</i>	1006	<i>Nitzschia</i> sp. I
936	<i>Nitzschia bilobata</i> var. <i>minor</i>	1007	<i>Berkeleya rutilans</i>
937	<i>Nitzschia pacifica</i>	1008	<i>Surirella gemma</i>
938	<i>Plagiogramma interruptum</i>	1009	<i>Hantzchia</i> sp.
939	<i>Pleurosigma acutum</i>	1010	<i>Aphanocapsa</i> sp.
940	<i>Pleurosigma delicatulum</i>	1011	<i>Coelosphaerium</i> sp.
941	<i>Pleurosigma rigidum</i>	1012	<i>Dactylococcopsis</i> sp.
942	<i>Synedra provincialis</i>	1013	<i>Eudorina</i> sp.
943	<i>Euastrum</i> sp.	1014	<i>Pandorina</i> sp.
944	<i>Navicula salinarum</i>	1015	<i>Coelastrum reticulatum</i>
945	<i>Lithodesmium undulatum</i>	1016	<i>Coelastrum</i> sp.
946	<i>Synedrosphenia gomphonema</i>	1017	<i>Micractinium pusillum</i>
947	<i>Navicula rhombica</i>	1018	<i>Melosira varians</i>
948	<i>Navicula laevissima</i>	1019	<i>Diploneis elliptica</i>
949	<i>Navicula hennedyii</i>	1020	<i>Achnanthes clevei</i>
950	<i>Lyngbya contorta</i>	1021	<i>Diploneis gruendleri</i>
951	<i>Navicula atomus</i>	1022	<i>Frustulia</i> sp.
952	<i>Navicula arvensis</i>	1023	<i>Rhabdonema</i> sp.
953	<i>Navicula irrorata</i>	1024	<i>Botryoccus protuberans</i>
954	<i>Navicula amphipleuroides</i>	1025	<i>Cosmarium costatum</i>
955	<i>Cyclotella</i> spp. > 30 um	1026	<i>Cosmarium</i> sp.
956	<i>Cyclotella caspia</i>	1027	<i>Dictyosphaerium planctonicum</i>
957	<i>Epithemia argus</i>	1028	<i>Dictyosphaerium pulchellum</i>
958	<i>Eunotia praerupta</i>	1029	<i>Kirchneriella obesa</i> major
959	<i>Auricula insecta</i>	1030	<i>Micrasterias</i> sp.
960	<i>Surirella striatula</i>	1031	<i>Pediastrum boryanum</i>
961	<i>Surirella fastuosa</i>	1032	<i>Pediastrum duplex</i> var. <i>rotundatum</i>
962	<i>Rhabdonema minutum</i>	1033	<i>Scenedesmus bernardii</i>
963	<i>Pinnularia major</i>	1034	<i>Scenedesmus hystrix</i>
964	<i>Stauroneis anceps</i> var. <i>hyalina</i>	1035	<i>Tetraedron lobulatum</i>
965	<i>Nitzschia amphibia</i>	1036	<i>Anabaena confervoides</i>
967	<i>Nitzschia proxima</i>	1037	<i>Chroococcus dispersus</i>
968	<i>Nitzschia vermicularis</i>	1038	<i>Lyngbya limnetica</i>
969	<i>Caloneis westii</i>	1039	<i>Merismopedia glauca</i>
970	<i>Cymbella turgidula</i>	1040	<i>Microcystis incerta</i>
971	<i>Gyrosigma balticum</i>	1041	<i>Oscillatoria tenuis</i>
972	<i>Proboscia inermis</i>	1042	<i>Hemiselmis</i> sp.
973	<i>Phormidium</i> sp.	1043	<i>Nephrochloris salina</i>
974	<i>Neidium affine</i>	1044	<i>Olisthodiscus</i> sp.
975	<i>Cymbella</i> sp.	1045	<i>Frustulia rhomboides</i>
976	<i>Mastogloia apiculata</i>	1046	<i>Merismopedia elegans</i> v. <i>major</i>

1047	Lyngbya sp.	1119	Tetraedron regulare var. incus
1048	Nephrochloris sp.	1120	Marssoniella elegans
1049	Schizothrix sp.	1121	Crucigenia lauterbornii
1050	Rhabdoderma sp.	1122	Errerella bornhemiensis
1051	Cyclotella spp.	1123	Chrysococcus tessellatus
1052	Cyclotella spp. 10 - 30 um	1124	Botryococcus sp.
1053	Rhabdoderma lineare	1125	Polydrieopsis spinulosa
1054	Dactylococcopsis fascicularis	1126	Ankistrodesmus spiralis
1055	Raphidiopsis curvata	1127	Pediastrum tetras
1056	Rhabdoderma sigmoidea f. minor	1128	Panadorina morum
1057	Amphiprora costata	1129	Microspora sp
1058	Staurastrum grande	1130	Microphytoflagellate sp #2 (unid)
1059	Nodularia sp.	1131	Aulacoseira herzogii
1060	Tetraedron regulare	1132	Ceratium carolinianum
1061	Gyrodinium aureolum	1133	Ceratium inflatum
1062	Gonium sp.	1134	Merismopedia convoluta
1063	Campylosira sp.	1135	Chlorophycean microphytoflagellates (unid.)
1064	Tetraedron gracile	1136	Euglena elastica
1065	Scenedesmus arcuatus	1137	Lagerheimia ciliata
1066	Mougeotia sp.	1138	Kephyrion sp.
1067	Tetraedron pentaedricum	1139	Caloneis lepidula
1068	Lagerheimia citrifomis	1140	Apedinella radians
1069	Lagerheimia sp.	1141	Closteriopsis longissima
1070	Chroococcus limneticus var elegans	1142	Pyrodinium bahamense
1071	Quadrigula lacustris	1143	Coelastrum cambricum
1072	Chroococcus prescottii	1144	Euglena spirogyra
1073	Desmids (Unid.)	1145	Pediastrum simplex var duodenarium
1074	Pleodorina sp.	1146	Scenedesmus perforatus
1075	Spirogyra sp.	1147	Phacus suecicus
1076	Chlorophycean filaments	1149	Trachelomonas bulla
1077	Dinobryon cylindricum	1150	Tetrademus smithii
1078	Rhizosolenia eriensis	1151	Spondylosium pygmaeum
1080	Glenodinium armatum	1152	Nephrocytium agardhianum
1081	Xanthidinium subhastiferum v. towerii	1153	Tetraedron limneticum
1082	Pleurotaenium tridentulum	1154	Eunotia serra var. diadema
1083	Micrasterias johnsonii	1155	Synura uvella
1084	Lagerheima longiseta	1156	Micrasterias truncata
1085	Oxyrrhis marina	1157	Dinobryon calciformis
1086	Pleurotaenium sp.	1158	Dinobryon bavaricum
1087	Quadrigula sp.	1159	Rhabdomonas spiralis
1088	Aphanothece sp.	1160	Staurastrum paradoxum var. cingulum
1089	Cryptomonad #1	1161	Licmophora tincta
1090	Micrasterias radiata	1162	Zygnema sp.
1091	Hyalotheca sp.	1163	Rhoicosphenia abbreviata
1092	Chrysophycean microphytoflagellates (unid)	1164	Lyngbya hieronymusii
1093	Kephyrion ovale	1165	Caloneis sp
1094	Arthrodesmus octocornis	1166	Euastrum abruptum
1095	Desmidium grevillii	1167	Navicula peregrina
1096	Elakatothrix gelatinosa	1168	Navicula cincta
1098	Protoperidinium wisconsinense	1169	Navicula placenta
1099	Gomphonema acuminatum	1170	Navicula eidrigeana
1100	Tetrastrum sp.	1171	Navicula spectabilis
1101	Selenastrum sp.	1172	Pinnularia legumen
1102	Scenedesmus denticulatus	1173	Pinnularia gibba
1103	Dictyosphaerium sp.	1174	Amphora exigua
1104	Euglenoid (Unid.)	1175	Cocconeis pediculus
1105	Ceratium hirundinella form brachyceras	1176	Suriella spiralis
1106	Gonatozygon brebissonii	1177	Tabellaria flocculosa
1107	Franceia sp.	1178	Bellerochea malleus
1108	Spondylosium planum	1179	Oocystis parva
1109	Actinastrum hantzschii var. fluviale	1180	Oocystis elliptica
1110	Desmidium baileyi	1181	Botryococcus sudeticus
1111	Cosmarium alpestre	1182	Pediastrum duplex gracilimum
1112	Chaetosphaeridium globosum	1183	Closterium archerianum
1113	Pleurotaenium nodulosum	1184	Chroococcus dispersus minor
1114	Volvox tertius	1185	Aphanothece gelatinosa
1115	Protoperidinium cinctum	1186	Gloeotheca linearis composita
1116	Glenodinium quadrifidum	1187	Phacus orbicularis
1117	Xanthidinium sp.	1188	Actinocyclus normanii f. normanii
1118	Micrasterias pinnatifida	1189	Stephanodiscus hantzschii

1190	<i>Cyclostephanos dubius</i>	1262	<i>Gloeocapsa aeruginosa</i>
1191	<i>Ulothrix</i> sp.	1263	<i>Gloeotheca</i> sp.
1192	<i>Protoperdinium aciculiferum</i>	1264	<i>Oscillatoria lacustris</i>
1193	<i>Gloeocapsa</i> sp.	1265	<i>Oscillatoria limosa</i>
1194	<i>Tetraedron hastatum</i>	1267	<i>Eunotia pectinalis</i>
1195	<i>Epithemia</i> sp.	1268	<i>Nitzschia acicularis</i>
1196	<i>Nephrocitium limneticum</i>	1269	<i>Surirella patella</i>
1197	<i>Odontella rhombus</i>	1270	<i>Centrtractus belanophorus</i>
1198	<i>Rhopalodia gibba</i>	1271	<i>Chrysococcus rufescens</i>
1200	<i>Closterium diana</i>	1272	<i>Dinobryon divergens</i>
1201	<i>Cosmarium turpinii</i>	1273	<i>Dinobryon sertularia</i>
1202	<i>Cochlodinium heterolobatum</i>	1274	<i>Mallomonas caudata</i>
1203	<i>Cochlodinium brandtii</i>	1275	<i>Mallomonas producta</i>
1204	<i>Botryococcus braunii</i>	1276	<i>Mallomonas tonsurata</i>
1205	<i>Gymnodinium neglectum</i>	1277	<i>Trachelomonas scabra</i> var. <i>longicollis</i>
1206	<i>Palmodictyon varium</i>	1278	<i>Ophiocytium capitatum</i> var. <i>longispinum</i>
1207	<i>Gymnodinium boguensis</i>	1279	<i>Synura adamsii</i>
1208	<i>Pleurosigma salinarum</i>	1280	<i>Euglena mutabilis</i>
1209	<i>Gymnodinium verruculosum</i>	1281	<i>Euglena viridis</i>
1210	<i>Selenastrum minutum</i>	1282	<i>Leptocinclis ovum</i> var. <i>gracilicauda</i>
1211	<i>Gymnodinium gracilentum</i>	1283	<i>Phacus latus</i>
1212	<i>Pavlova homersandii</i>	1284	<i>Phacus monilatus</i>
1213	<i>Oscillatoria subbrevis</i>	1285	<i>Phacus perkinensis</i>
1214	<i>Isochrysis galbana</i>	1286	<i>Phacus triqueter</i>
1215	<i>Aphanizomenon</i> sp.	1287	<i>Strombomonas australica</i>
1216	<i>Acanthosphaera zachariasii</i>	1288	<i>Trachelomonas acanthophora</i>
1217	<i>Actinastrum hantzschii</i> var. <i>elongatum</i>	1289	<i>Trachelomonas armata</i> var. <i>longa</i>
1219	<i>Ankistrodesmus falcatus</i> var. <i>mirabilis</i>	1290	<i>Trachelomonas globularis</i> var. <i>boyeri</i>
1220	<i>Asterococcus limneticus</i>	1291	<i>Trachelomonas hispida</i> var. <i>coronata</i>
1221	<i>Coelastrum microporum</i>	1292	<i>Trachelomonas planctonica</i> var. <i>oblonga</i>
1222	<i>Eudorina elegans</i>	1293	<i>Trachelomonas raciborskii</i>
1223	<i>Franceia ovalis</i>	1294	<i>Trachelomonas regulosa</i>
1224	<i>Gonium pectorale</i>	1295	<i>Trachelomonas similis</i>
1225	<i>Kirchneriella elongata</i>	1296	<i>Trachelomonas superba</i>
1226	<i>Kirchneriella subsolitaria</i>	1297	<i>Trachelomonas superba</i> var. <i>duplex</i>
1227	<i>Micractinium pusillum</i> var. <i>elegans</i>	1298	<i>Trachelomonas varians</i>
1228	<i>Pleurocapsa minor</i>	1299	<i>Trachelomonas verrucosa</i>
1229	<i>Quadrigula chodatii</i>	1300	<i>Trachelomonas volvocina</i>
1230	<i>Rhizochrysis limnetica</i>	1301	<i>Gymnodinium fusum</i>
1231	<i>Scenedesmus incrassatulus</i>	1302	<i>Peridinium aciculiferum</i>
1232	<i>Selenastrum gracile</i>	1303	<i>Peridinium cinctum</i>
1233	<i>Selenastrum westii</i>	1304	<i>Peridinium</i> sp.
1234	<i>Spirogyra crassa</i>	1305	<i>Cryptomonas erosa</i>
1235	<i>Quadrigula closterioides</i>	1306	<i>Cryptomonas erosa</i> var. <i>reflexa</i>
1236	<i>Scenedesmus obliquus</i>	1307	<i>Cryptomonas massonii</i>
1237	<i>Spirogyra tenuissima</i>	1308	<i>Cryptomonas ovata</i>
1238	<i>Tetraedron arthrodesmiforme</i>	1309	<i>Cryptomonas phaseolus</i>
1239	<i>Tetraedron cruciatum</i>	1310	<i>Cryptomonas rostrata</i>
1240	<i>Tetraedron regulare</i> var. <i>torsum</i>	1311	<i>Rhodomonas minuta</i>
1241	<i>Volvox aureus</i>	1312	<i>Dinobryon sociale</i>
1242	<i>Arthrodesmus incus</i> var. <i>extensus</i>	1313	<i>Gleocapsa punctata</i>
1243	<i>Closterium limeatum</i>	1314	<i>Pediastrum tetras</i> var. <i>tetraodon</i>
1244	<i>Closterium pronum</i>	1315	<i>Cladophora</i> sp.
1245	<i>Closterium setaecum</i>	1316	<i>Pediastrum obtusum</i>
1246	<i>Cosmarium cynthia</i>	1317	<i>Euglena polymorpha</i>
1247	<i>Penium</i> sp.	1318	<i>Euglena convoluta</i>
1248	<i>Pleurotaenium trabecula</i>	1319	<i>Cosmarium contractum</i>
1249	<i>Staurastrum cingulum</i> var. <i>floridense</i>	1320	<i>Zygabikodinium lenticulatum</i>
1250	<i>Staurastrum leptocladum</i> var. <i>cornutum</i>	1321	<i>Peridinium pseudolaeva</i>
1251	<i>Staurastrum pentacerum</i>	1322	<i>Gymnodinium cnecoides</i>
1252	<i>Anabaena affinis</i>	1323	<i>Protoperdinium decipiens</i>
1253	<i>Anabaena aequalis</i>	1324	<i>Peridinium lomnickii</i> var. <i>splendida</i>
1254	<i>Anabaena augstumalis</i> var. <i>marchica</i>	1325	<i>Aulacoseira</i> sp.
1255	<i>Anabaena limnetica</i>	1326	<i>Aulacoseira granulata</i> var. <i>curvata</i>
1256	<i>Anabaena circinalis</i>	1327	<i>Cymbella affinis</i>
1257	<i>Anabaena spiroides</i> var. <i>crassa</i>	1328	<i>Caloneis trinodis</i>
1258	<i>Anabaena wisconsinense</i>	1329	<i>Cosmarium subreniforme</i>
1259	<i>Aphanocapsa delicatissima</i>	1330	<i>Cosmarium tenue</i>
1260	<i>Aphanocapsa grevillei</i>	1331	<i>Closterium parvulum</i>
1261	<i>Aphanocapsa pulchra</i>	1332	<i>Arthrodesmus validus</i> var. <i>incrassatus</i>

1333	Gymnodinium uberrimum	1363	Pennate diatoms < 10-20um
1334	Gymnodinium mitratum	1364	Cryptomonas sp. < 10um
1335	Peridinium inconspicuum	1365	Aphanizomenon flos-aqua
1336	Cocconeis flumiatilis	1366	Thalassiosira anguste-lineata
1337	Gymnodinium impatiens	1367	Unid. Centric Diatom Diam 10-30 um
1338	Cosmarium ornatum	1368	Unid. Centric Diatom Diam 31-60 um
1339	Cylindrospermum doryphorum	1369	Unid. Centric Diatom Diam >60 um
1340	Hyalotheca dissiliens var. tatica	1370	Unid. Pennate Diatom Length 10-30 um
1341	Spirulina laxa	1371	Unid. Pennate Diatom Length 31-60 um
1342	Spirulina major	1372	Unid. Pennate Diatom Length >60 um
1343	Hydrodictyon reticulatum	1373	Caloneis lewisii
1344	Treubaria setigerum	1374	Encyonema silesiacum
1345	epiphytic flagellates (unid)	1375	Navicula elegans
1346	Microcoleus sp.	1376	Navicula phyllepta
1347	Stigeoclonium glomeratum	1377	Navicula lyra
1349	Blue Green Trichome B (unid)	1378	Navicula spicula
1350	Lepocinclis sphagnophila	1379	Suriella Splendida
1351	Carteria cordiformis	1380	Surirella Crumena
1352	Microspora quadrata	1381	Tabularia fasciculate
1353	Hormidium Klebsii	1382	Hydrodictyon
1354	Oocystis Borgei	1383	Nitzschia frustulum perpulsilla
1355	Pleurotaenium subcoronulatum var. detum	1384	Scenedesmus opoliensis
1356	Peridinium gatunense	1385	Scenedesmus brasiliensis
1357	Dictyocha Siderea	1386	Guinardia striata
1358	Navicula septentrionalis	1387	Aphanizomenon issatschenkoii
1359	Characium limneticum	1388	Blue Green Trichome cell
1360	Centric diatom < 10um	1389	Blue Green Trichome B cell
1361	Centric diatom 10-20um	1390	Alexandrium monilatum
1362	Pennate diatoms < 10um		

## #VARIABLE NAMES AND DESCRIPTIONS FOR DATA FILES

Structure for data files on: <http://www.chesapeakebay.net/>

## &gt;PHYTOPLANKTON OR PICOPLANKTON TAXONOMIC ABUNDANCE AND COMPOSITON FILES

Name	Type	Width	Variable Description
SOURCE	Text	10	Data Collection Agency
SAMPLE_TYPE	Text	2	Sample Collection Type
CRUISE	Text	6	Chesapeake Bay Program Cruise Number
STATION	Text	15	Sampling Station
SAMPLE_DATE	Date/Time	8	Sampling Date (YYYYMMDD)
LAYER	Text	3	Layer of Water Column in Which SampleWas Taken
SAMPLE_NUMBER	Number	4	Sample Replicate Number
GMETHOD	Text	3	Chesapeake Bay Program Gear Method Code
TSN	Text	7	ITIS Taxon Serial Number
LATIN_NAME	Text	45	Species Latin Name
SIZE	Text	30	Cell Size Groupings when taken
METHOD	Text	8	Chesapeake Bay Program Sample Analysis Code
PARAMETER	Text	15	Sampling Parameter Name
VALUE	Number	8	Sampling Parameter Value
UNITS	Text	15	Sampling Parameter Reporting Units
NODCCODE	Text	12	National Oceanographic Data Center Species Code
SPEC_CODE	Text	14	In House Species Code
SER_NUM	Text	12	Sample Serial Number
R_DATE	Date/Time	8	Version Date of Data (YYYYMMDD)

## &gt; PHYTOPLANKTON AND PICOPLANKTON SAMPLING EVENT FILES

Name	Type	Width	Variable Description
SOURCE	Text	10	Data Collection Agency
SAMPLE_TYPE	Text	2	Collection Type
CRUISE	Text	6	Chesapeake Bay Program cruise number
SAMPLE_DATE	Date/Time	8	Sampling date (YYYYMMDD)
LATITUDE	Number	8	Latitude in decimal degrees



LONGITUDE	Number	8	Longitude in decimal degrees
P_DEPTH	Number	4	Composite Sample Cut Off Depth
R_DATE	Date/Time	8	Data version date (YYYYMMDD)
SALZONE	Text	2	Salinity zone
SAMPLE_VOLUME	Number	8	Total Volume of Sample
UNITS	Text	15	Reporting Units of Sample Volume
STATION	Text	15	Sampling Station
TOTAL_DEPTH	Number	4	Total Station Depth (meters)
SAMPLE_TIME	Date/Time	8	Sample Collection Time (HHMM)

>The following field may also appear in a downloaded data set:

Name	Type	Width	Variable Definitions
BASIN	Text	20	Chesapeake Bay Basin Designation
HUC8	Text	8	USGS Eight Digit Hydrologic Unit Code
CATALOGING_UNIT_DESCRIPTION	Text	50	USGS Cataloging Unit Code Description
FIPS	Text	5	Federal Information Processing Code
STATE	Text	3	Federal Information Processing Code State Designation
COUNTY_CITY	Text	30	Federal Information Processing Code City or County Designation
LL_DATUM	Text	5	Latitude and Longitude Geographic Datum
CBSEG_1998	Text	6	1998 Chesapeake Bay Segment Designation
CBSEG_1998_DESCRIPTION	Text	50	1998 Chesapeake Bay Segment Designation Description

#### # VARIABLES NAMES AND DESCRIPTIONS FOR SPECIES KEY

##### > PHYTOPLANKTON AND PICOPLANKTON SPECIES KEY

File of name format: VAPHKYyy.TXT

Name	Type	Width	Variable Descriptions
SPEC_CODE	Text	14	Source In-House Species Codes
SOURCE	Text	6	Data Source Identifier
DATA_TYPE	Text	2	Data Type Identifier Code
SOURCE_LBL	Text	45	Source Species Latin Name
LBL	Text	45	National Oceanographic Data Center Species Latin Name
TSN	Text	7	ITIS Taxon Serial Number
R_DATE	Date/Time	8	Version Date of Data (YYYYMMDD)
VOLUME	Number	8	Cell Biomass Estimator
SIZE	Text	30	Taxa Size-Fraction Identifier
LIFE_STG	Text	3	Chesapeake Bay Program Life Stage Code

#### # REFERENCE CODES IN DATA FILES AND TAXONOMIC KEY

See 2000 Users Guide to Chesapeake Bay Program Biological and Living Resources Monitoring Data for full listing.

>MISSING VALUES: Missing Sampling Times have been replaced with 00:00.

> DATA\_TYPE: Data Type

BE Benthic  
 FL Fluorescence  
 MI Microzooplankton  
 MZ Mesozooplankton  
 PD Primary Production  
 PH Phytoplankton  
 PP Picoplankton

>SOURCE: Data Collection Agency  
 ODU - Old Dominion University

>SAMPLE\_TYPE: Collection Type  
 C - Field Composite Sample, sample composed of subsamples from multiple depths

>CRUISE: Chesapeake Bay Program Cruise Number  
 See 2000 USERS GUIDE for complete listing of CBP cruise numbers

>GMETHOD: Sampling Gear Code  
 07 - unspecified plankton pump

>LAYER: Layer of Water Column in which Sample was Taken  
 AP- Above Pycnocline  
 BP- Below Pycnocline  
 WC- Whole Water Column

>NODCCODE: National Oceanographic Data Center Species Code, Version 8.  
 NOTE: For current listing of Chesapeake Bay species and their codes, see The 1997 Chesapeake Bay Basin Species list. Organisms with out current NODC Codes have been assigned partial NODC codes containing alphabetic where no code has been assigned.

>SALZONE: Salinity zone  
 F - Tidal fresh (0 - 0.5 ppt)  
 O - Oligohaline (0.5 - 5.0 ppt)  
 M - Mesohaline (5.0 - 18.0 ppt)  
 P - Polyhaline (>18.0 ppt)  
 \*E- An F,O,M, or P followed by an E indicate an estimated salinity range based on salinity data collected within a week of the biological sampling event. Used only when no actual salinity data available.

>BASIN: Chesapeake Bay Program Basin Designations  
 BAY- Chesapeake Bay  
 ELZ- Elizabeth River  
 JAM- James River  
 YRK- York River  
 RAP- Rappahanock River

>TSN: IT IS Taxon Serial Numbers  
 Note for current listing of Chesapeake Bay Program Species and their codes; see 1998 Bay Basins Species List. Organisms without current serial numbers have ALL been assigned TSN of BAYXXXX

> CBSEG\_1998: Chesapeake Bay Program Monitoring Segment

CBSEG_1998	DESCRIPTION
CB6PH	CHESAPEAKE BAY-POLYHALINE REGION
CB7PH	CHESAPEAKE BAY-POLYHALINE REGION
CB8PH	CHESAPEAKE BAY-POLYHALINE REGION
JMSOH	JAMES RIVER-OLIGOHALINE REGION
JMSPH	JAMES RIVER-POLYHALINE REGION
JMSTF	JAMES RIVER-TIDAL FRESH REGION
MOBPH	MOBJACK BAY-POLYHALINE REGION
PMKOH	PAMUNKEY RIVER-OLIGOHALINE REGION
PMKTF	PAMUNKEY RIVER-TIDAL FRESH REGION

CBSEG_1998	DESCRIPTION
RPPMH	RAPPAHANNOCK RIVER-MESOHALINE REGION
RPOH	RAPPAHANNOCK RIVER-OLIGOHALINE REGION
SBEMH	SOUTH BRANCH ELIZABETH RIVER-MESOHALINE REGION
YRKMH	YORK RIVER-MESOHALINE REGION

>FIPS: Federal Information Processing Codes

FIPS	STATE	COUNTY
51095	VA	JAMES CITY
51097	VA	KING AND QUEEN
51103	VA	LANCASTER
51127	VA	NEW KENT
51131	VA	NORTHAMPTON
51149	VA	PRINCE GEORGE
51159	VA	RICHMOND
51199	VA	YORK
51550	VA	CHESAPEAKE CITY
51650	VA	HAMPTON
51740	VA	PORTSMOUTH
51810	VA	VIRGINIA BEACH

>HUC8: USGS Hydrologic Unit Codes

HUC8	CATALOGING_UNIT_DESCRIPTION
02050306	LOWER SUSQUEHANNA
02080101	LOWER CHESAPEAKE BAY
02080104	LOWER RAPPAHANNOCK
02080106	PAMUNKEY
02080107	YORK
02080206	LOWER JAMES
02080208	HAMPTON ROADS

>METHOD: Chesapeake Bay Program Lab Method Code Designation

PP101  
PH102  
PH103

>PARAMETER and UNIT: Measured Parameter and reporting units.

PARAMETER	UNITS
COUNT	NUMBER/LITER

# NUMERIC VARIABLE WARNING AND ERROR BOUNDS

Variable	Valid Range
SAMPLE_DATE	19850101-20031231
COUNT	3172 - 102224636
MAXDEPTH	0.5 - 32.0
R_DATE	19950301-20041231
SAMPLE_NUMBER	1-2
LATITUDE	See Station Names, Latitudes, Longitudes and Depths.
LONGITUDE	See Station Names, Latitudes, Longitudes and Depths.
P_DEPTH	0.5-<TDEPTH Note this is a composite cut off not pycnocline depth!

R_DATE	19950301-19991230
SAMVOL_L	12-200
TOTAL_DEPTH	1.8-33
SAMPLE_TIME	0651-1935, 00:00 INDICATES MISSING TIME

#### # IMPORTANT DATA REVISIONS

THE LIVING RESOURCES DATA MANAGER RECOMMENDS THAT ALL DATA ANALYSIS BE PERFORMED WITH THE MOST RECENT DATA SETS VERSIONS AVAILABLE. HOWEVER IF YOU HAVE BEEN WORKING WITH OLDER DATA SETS THE FOLLOWING ARE IMPORTANT CHANGES TO BE AWARE OF.

CELL BIOMASS CAN BE ESTIMATED FROM THE CELL VOLUMES PROVIDED IN THE TAXON KEY FILE. THESE MEASUREMENTS ARE THE AVERAGE CELL VOLUMES BASED ON MEASUREMENT MADE BY THE PRINCIPLE INVESTIGATORS.

6/30/1996- All plankton data was resubmitted to the Chesapeake Bay Program office due to discrepancies in sampling dates between synchronously collected samples. Sampling dates were corrected to field logs and resubmitted to the Data Center. Please do not use data with an R\_DATE prior to 06/01/96.

8/31/1995- GMETHOD was changed to 7. Code 7 refers to an unspecified plankton pump. For an extensive gear code list see Table 17, PAGE F-9 APPENDIX F, of the Living Resources Data management plan, 1989. This is a change from GMETHOD code in previous versions of the data set. This does not represent a change in actual sampling gear.

8/31/1993- LBL all Latin Names and spelling for names have been corrected to the National Oceanographic Data Center accepted spelling.

8/31/1995- CRUISE NUMBERS - BAY012-BAY211 were supplied by the Chesapeake Bay Program office. See the Guide to Living Resources Data Sets for complete listing of Cruise periods.

8/31/1995- SER\_NUM Old Dominion University does not use a serial number system for phytoplankton sample tracking so this variable is not available

8/31/1995- P\_DEPTH >0.5-<TDEPTH Note this is a composite sample cut off depth. This depth is not the pycnocline depth!

SUMMER 1997 - The Living Resources Data manager supplied salinity zones to the plankton data based on salinity data collected by the Virginia Water Quality Monitoring Program. Values were derived from Water Quality Hydrographic data collected concurrently with the mesozooplankton. If data was not available for the of sampling but was collected within a one week window of sampling date, the water quality data was used to determine a salinity zone. However the salinity zone is marked with an E to denote being estimated.

02/01/1998- The salinity zones appearing in the 1997 data are provisional. They have not yet been checked against the water quality data for validation. The 1997 Virginia Tributary water quality data will not be delivered to the CBPO until June 1998. After delivery of the water quality data, salinity zones will be confirmed. Salinity zones will be filled in when the corresponding Water Quality monitoring data becomes available.

01/01/1999- Due to the 1998 CBP Living Resources split sampling program it was determined that there was a nomenclature difference between laboratories in Maryland and Virginia. The species Merismopedia (VA species name) and Agmenellum (MD species name) were determined to be synonymous. After a literature review both states agreed to use the genera designation Merismopedia. Please contact the Living resources data manager for details.

01/01/2000- All Latitudes and Longitudes converted to NAD83 coordinates.

August 2002: ODU purchased new inverted plankton microscopes changing their mid- and high magnification from 315/500X to 300/600X, with magnification constants adjusted accordingly.

Winter 2002- For extensive details in regards to quality assurance issues and data comparability issues between Maryland and Virginia Programs please see the CBP Phytoplankton Split sample portion of the Chesapeake Bay Quality Assurance Program at:

<http://www.chesapeakebay.net/qualityassurance.htm>

08/11/2005. Note due to contract changes starting in January 1996, station LE5.5 had a coordinate change. This station move was not documented until August 2005. Due to this station relocation, all data collected at the altered location had the station name changed to LE5.5-W in August 2005.

01/01/2005- All data enumerated using new uniform bay wide counting technique. There will be a significant increase in the number of taxa identified in Maryland samples counted after 1/1/2005. Please be aware of this potential source of step trend in the data.

04/14/2006-Missing Data Report for July-Dec 2005 Data. 1) October 2005- Phytoplankton and picoplankton samples not collected at station CB6.4 due to inclement weather. 2) December 2005- Phytoplankton and picoplankton samples not collected at stations CB6.1, CB7.3e, CB7.4, LE3.6 due to boat malfunctions. 3) August 2005- Phytoplankton sample WE4.2 BP sample leaked. Autotrophic picoplankton count: 324,099,200 cells/liter. 4) October 2005- Picoplankton sample TF4.2 sample leaked.

11/04/2008- In March 2008, CB6.1, CB6.4, LE3.6, and WE4.2 were not collected due to weather. This means that the above and below phytoplankton and picoplankton samples, as well as the productivity were not collected. Also the March LE5.5W was collected on April 1, 2008, due to rescheduling from the same foul weather. During the RET5.2 collection in May, there was a problem with the sampling pump, and the BP samples could not be collected.

10/18/2010- . During the period from January–June 2010 there were no collections made in January or February due to budget/contract issues. There were also four stations that were not collected in May. TF4.2 was not collected due to boat problems, RET4.3 was not collected due to staffing/scheduling problems, and CB6.1 and LE3.6 were not collected due to foul weather.

01/03/2011- From July 2010 on wardt here will be no BP collections made in the tributary stations (RET3.1, RET4.3, RET5.2, SBE5, TF3.3, TF4.2, TF5.5). AP stations and collection protocols remain constant, as do samples collected at the BAY stations. There were also two stations that were not collected due to foul weather. RET4.3 was not collected in August, and RET5.2 was not collected in September.

# KEY WORDS (EXCLUDING VARIABLE NAMES)

Phytoplankton species  
Phytoplankton Counts  
Phytoplankton Densities  
Phytoplankton Monitoring

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**THIS IS THE END OF THE VIRGINIA CHESAPEAKE BAY PROGRAM  
PHYTOPLANKTON DATA DICTIONARY**

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