

Oyster Best Management Practices (BMP) in the Chesapeake Bay: Management and Scientific Considerations

Fisheries GIT Meeting
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Acknowledgements

Role	Oyster BMP Expert Panel
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Chesapeake Bay Program/US EPA Advisors	<p>Lucinda Power, Rich Batiuk, Lew Linker, Matt Johnston, Ralph Spagnolo, Ed Ambrogio, Jeff Sweeney, Jeremy Hanson, Julie Reichert-Nguyen</p>
Panel Support	<p>Ward Slacum and Emily French (ORP), Carl Cerco (US Army Corps of Engineers), Paige Hobaugh, Emilie Franke, and Kyle Runion (CBP)</p>
Data	<p>Colleen Higgins, Kurt Stephenson, Bonnie Brown, Peter Kingsley-Smith, Steve Allen, Paige Ross, Roger Mann, and Melissa Southworth</p>
Stakeholder Review	<p>Citizen Advisory Committee, Chesapeake Bay Foundation, Chesapeake Bay Commission, Lynnhaven River Now, Southern Environmental Law Center and partners, Oyster Company of Virginia, and Norfolk Public Works</p>

Decision Framework: Incremental BMP Recommendations

Oyster Practice Category x Crediting Protocol	Private Oyster Aquaculture					Licensed Oyster Harvest				Oyster Reef Restoration		
	A. Off-bottom private oyster aquaculture using hatchery-produced oysters	B. On-bottom private oyster aquaculture using hatchery-produced oysters	C. On-bottom private oyster aquaculture using transplanted wild oysters	D. On-bottom private oyster aquaculture using substrate addition	E. Private oyster aquaculture with no activity	F. Licensed oyster harvest using hatchery-produced oysters	G. Licensed oyster harvest using transplanted wild oysters	H. Licensed oyster harvest using substrate addition	I. Licensed oyster harvest with no activity	J. Oyster reef restoration using hatchery-produced oysters	K. Oyster reef restoration using substrate addition	L. Oyster reef restoration using no harvest area designation only
1. Nitrogen Assimilation in Oyster Tissue	1 st Approved	1 st Approved	1 st Not Endorsed	1 st Approved	1 st Not Endorsed	2 nd Complete	2 nd Not Endorsed	Later	2 nd Not Endorsed	2 nd Complete	2 nd Complete	2 nd Policy Issue
2. Nitrogen Assimilation in Oyster Shell	2 nd Research Gap	2 nd Research Gap	2 nd Not Endorsed	2 nd Research Gap	2 nd Not Endorsed	2 nd Research Gap	2 nd Not Endorsed	Later	2 nd Not Endorsed	2 nd Complete	2 nd Complete	2 nd Policy Issue
3. Enhanced Denitrification Associated with Oysters	2 nd Research Gap	2 nd Research Gap	2 nd Not Endorsed	2 nd Research Gap	2 nd Not Endorsed	2 nd Research Gap	2 nd Not Endorsed	Later	2 nd Not Endorsed	2 nd Complete	2 nd Complete	2 nd Policy Issue
4. Phosphorus Assimilation in Oyster Tissue	1 st Approved	1 st Approved	1 st Not Endorsed	1 st Approved	1 st Not Endorsed	2 nd Complete	2 nd Not Endorsed	Later	2 nd Not Endorsed	2 nd Complete	2 nd Complete	2 nd Policy Issue
5. Phosphorus Assimilation in Oyster Shell	2 nd Research Gap	2 nd Research Gap	2 nd Not Endorsed	2 nd Research Gap	2 nd Not Endorsed	2 nd Not Endorsed	2 nd Not Endorsed	Later	2 nd Not Endorsed	2 nd Complete	2 nd Complete	2 nd Policy Issue
6. Suspended Sediment Reduction Associated with Oysters	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later
7. Enhanced Nitrogen Burial Associated with Oysters	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later
8. Enhanced Phosphorus Burial Associated with Oysters	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later	Later

1st Report (approved)

2nd Report (in draft)

Later = Future Report

Endorsed Licensed Oyster Harvest-Definition

Endorsed Oyster Practice-Protocol Combinations for BMP Approval

This chapter describes the Panel's recommendations on the nitrogen and phosphorus reduction effectiveness determination concerning assimilation in tissue of harvested oysters for the below oyster practice:

Endorsed Oyster Practice Title	Reduction Protocol	Practice Definition
F. Licensed oyster harvest using hatchery-produced oysters	1. Nitrogen assimilation in oyster tissue	Planting oysters (e.g., spat-on-shell, single oysters) produced from hatchery techniques directly on the bottom to enhance the stock in State-designated fishing areas (e.g., public shellfish fishing grounds) for eventual removal (harvest) from the water by individuals holding the proper licenses.
	4. Phosphorus assimilation in oyster tissue	

Licensed Oyster Harvest-Qualifying Conditions

- **Max Oyster Harvest Allowance for Reduction Credit**—Harvest claimed for reduction credit can be at maximum no more than 15% of hatchery-produced oyster spat planted per planting.
- **Oyster Harvest Allowance Timeframe**—Maximum of five years from the planting of hatchery-produced oysters.
- **Reduction Credit Time Lag**—Oyster harvest becomes eligible for reduction credit two years after a planting of hatchery-produced oysters (single oysters or spat-on-shell less than 2 inches) unless an assessment is done demonstrating a timeframe less than two years is appropriate.
- **Harvest Reduction Crediting Timeframe**—Oyster harvest allowance timeframe minus the reduction credit time lag equals the timeframe the nitrogen and phosphorus reduction from the oyster harvest allowance can be credited.
- **Lifespan of BMP**—Treatments (i.e., plantings) must occur within five years for the BMP to remain active. Inactivity will cause the BMP to expire until another treatment occurs.

Approved Oyster Reef Restoration Practices for Interim BMP Use (Pending Approval for Crediting)

Endorsed Oyster Practice-Protocol Combinations for BMP Approval

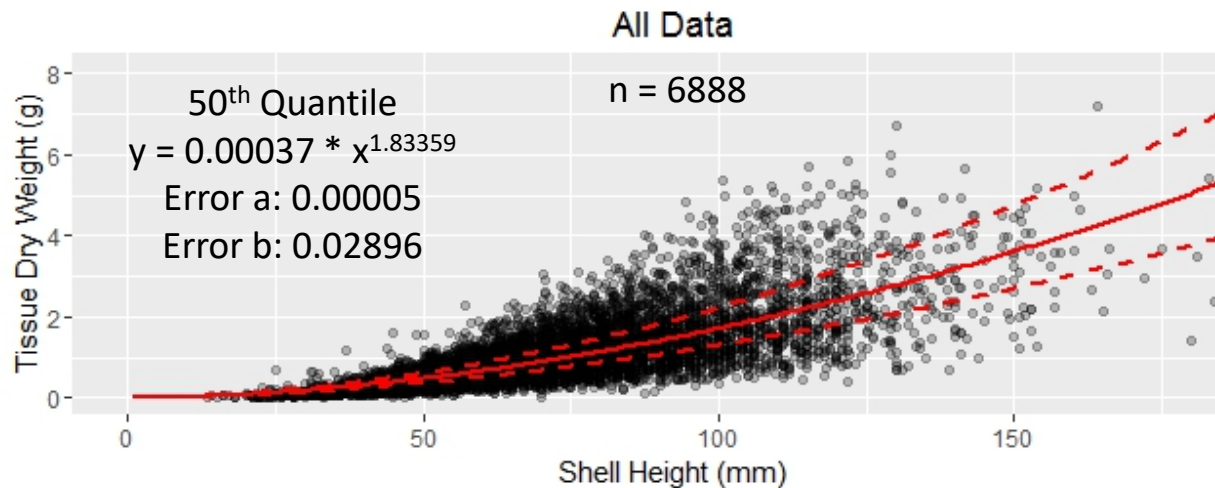
This chapter describes the Panel's recommendations on the nitrogen and phosphorus reduction effectiveness determination concerning assimilation (sequestration) in tissue and shell for the below oyster practices:

Endorsed Oyster Practice Title	Reduction Protocols	Practice Definition
J. Oyster reef restoration using hatchery-produced oysters	1. Nitrogen assimilation in oyster tissue 2. Nitrogen assimilation in oyster shell	Planting oysters (e.g., spat-on-shell, single oysters) produced from hatchery techniques directly on the bottom or raised substrate to enhance oyster biomass in areas where removal is not permitted.
K. Oyster reef restoration using substrate addition	4. Phosphorus assimilation in oyster tissue 5. Phosphorus assimilation in oyster shell	Planting oyster shells and/or alternative substrate directly on the bottom to attract recruitment of naturally occurring (wild) oyster larvae to enhance oyster biomass in areas where removal is not permitted.

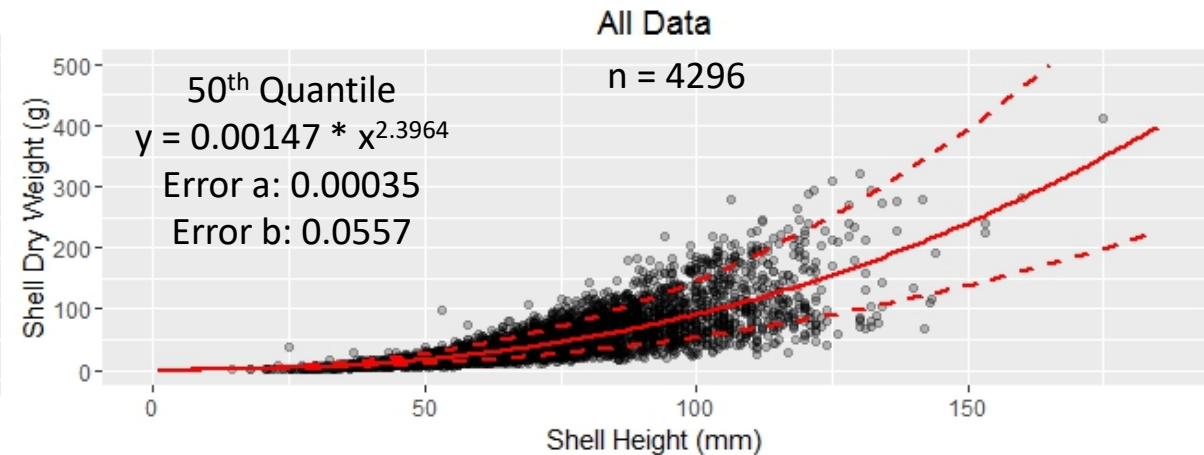
Oyster Reef Restoration: Calculating N & P Assimilation In Tissue & Shell

- Uses Chesapeake Bay-wide shell height to tissue and shell dry weight regression equations

Chesapeake Bay Oyster Tissue Reef Data



Chesapeake Bay Oyster Shell Reef Data



- Full dataset (Chesapeake Bay-Wide)
- 50th quantile full dataset
- - Error

- The average nitrogen (tissue: 8.2%, shell: 0.2%) and phosphorus (tissue: 0.9%, shell: 0.04%) percent contents were used to calculate the amount sequestered in the live oyster tissue and shell biomass (tissue percent contents approved by CBP, Cornwell et al. 2016; shell percent contents in draft).

N & P Assimilation Interim BMPs—Combined Tissue & Shell Based on 2017/2018 Harris Creek Data (Shell-Base Sites)

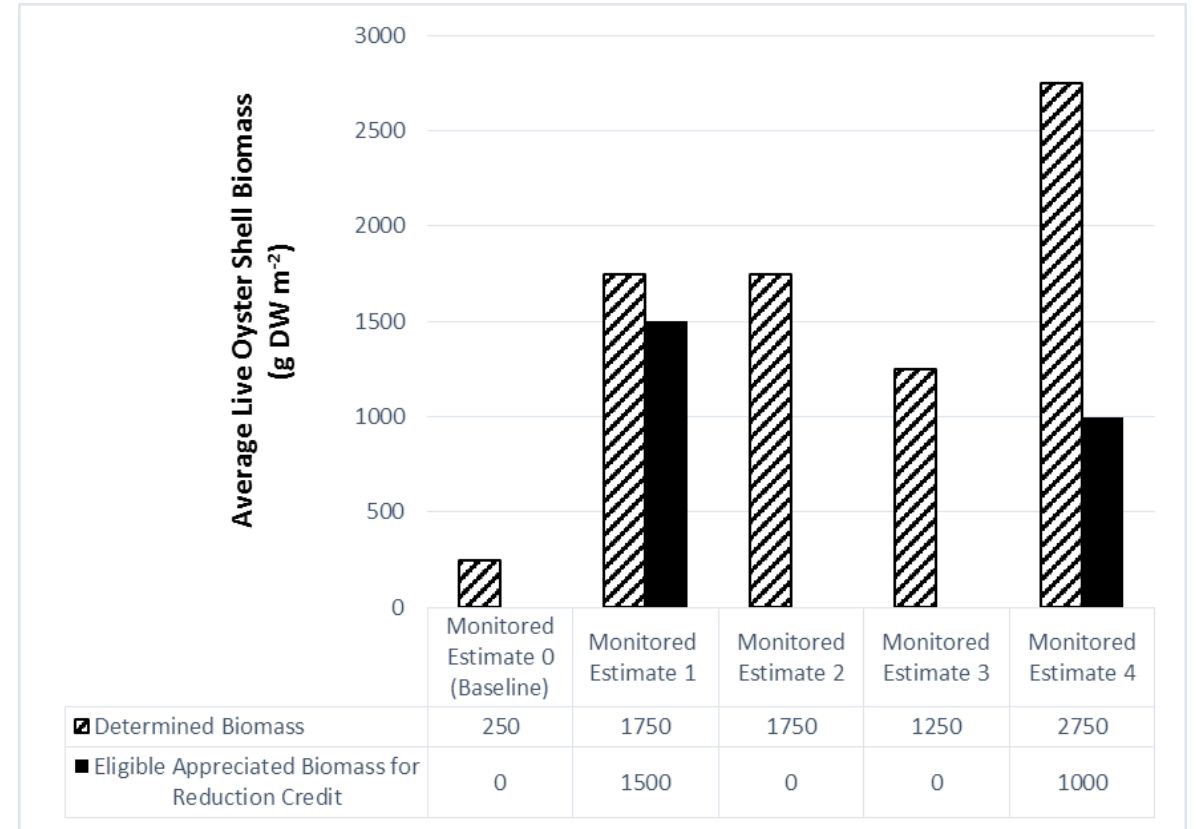
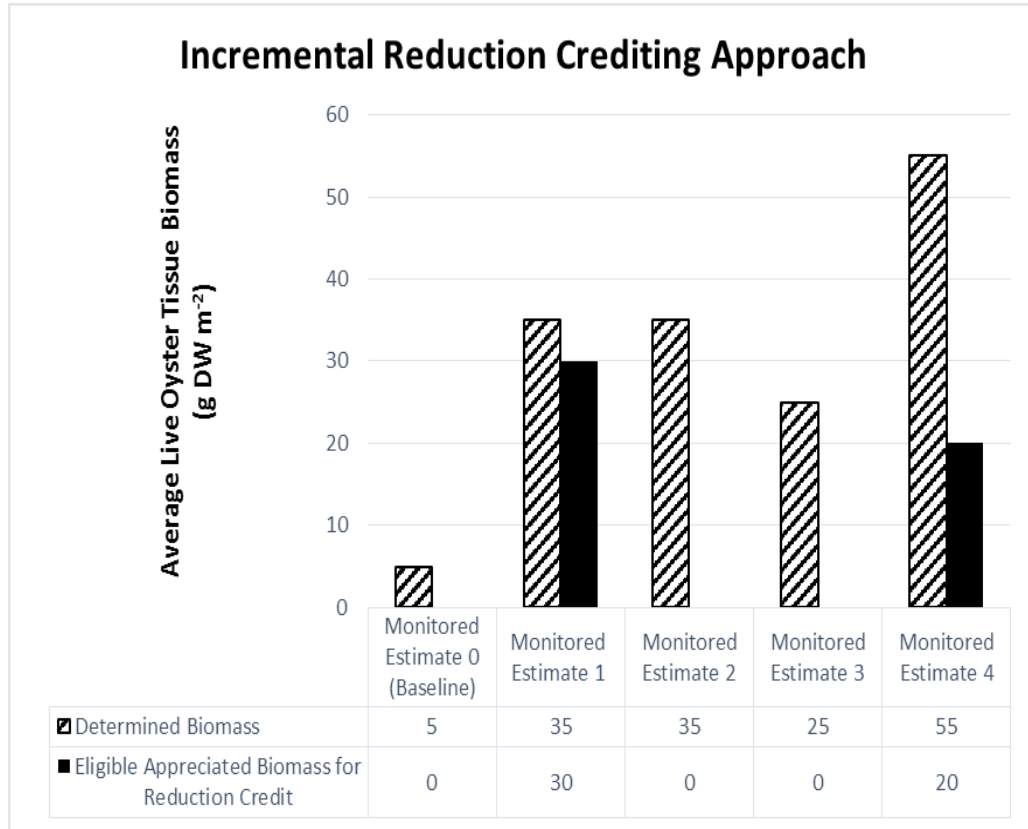
Oyster Reef Restoration-Assimilation Estimates for Planning		
Live Oyster Tissue + Shell Reduction Effectiveness	lbs acre⁻¹ year⁻¹ (max duration = 3 years)	
Treatment Category	Nitrogen	Phosphorus
Shell-Base + SOS n = 7 sites Mean Live Density = 42 oysters m⁻²	24	4

- Planning estimates are one time credits expressed as annual rates. Can be applied for a total of three years on acres where substrate (shell or alternative substrate, such as granite or stone) and/or hatchery-produced SOS were planted.
- The total N and P reduction (tissue plus shell) ~ 74 lbs acre⁻¹ and 12 lbs acre⁻¹, respectively.
- Represents a mean live oyster tissue and shell biomass of ~ 45 and 2300 g m⁻², respectively, three years post-restoration.

Site-Specific Oyster Reef Restoration BMP for Crediting: Assimilation Reduction Effectiveness Strategy

- **Type of reduction:** One-time credit that can be credited incrementally based on appreciated tissue and shell biomass from live oysters during the monitoring timeframe of at least every three years established by the implementing program.
- **Useful life of the BMP:** No reduction credit is given if monitored oyster biomass is below the established pre-restoration baseline value, does not increase from the previously credited post-restoration amount, or decreases.

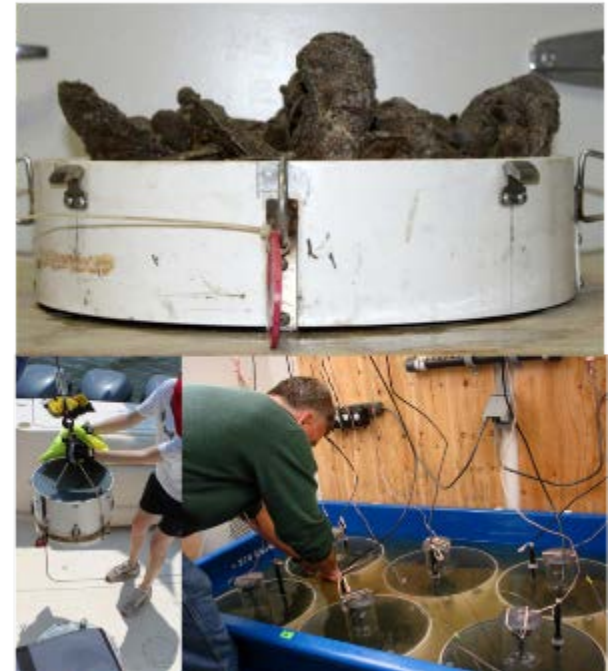
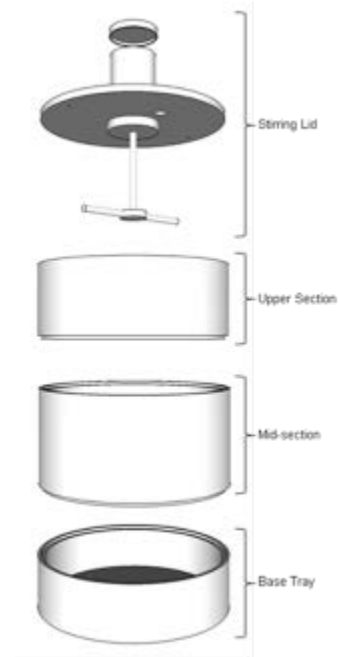
Site-Specific Oyster Reef Restoration BMP: Assimilation Reduction Effectiveness Strategy Example



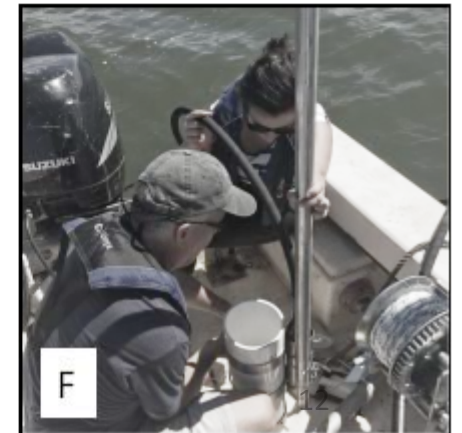
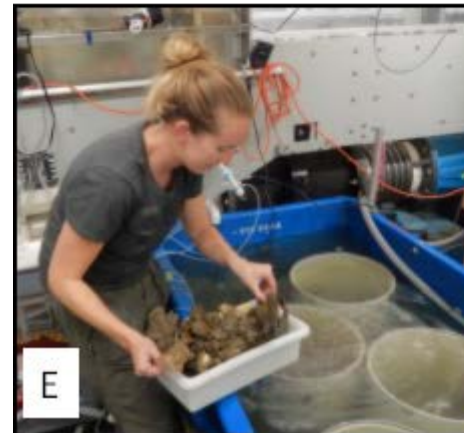
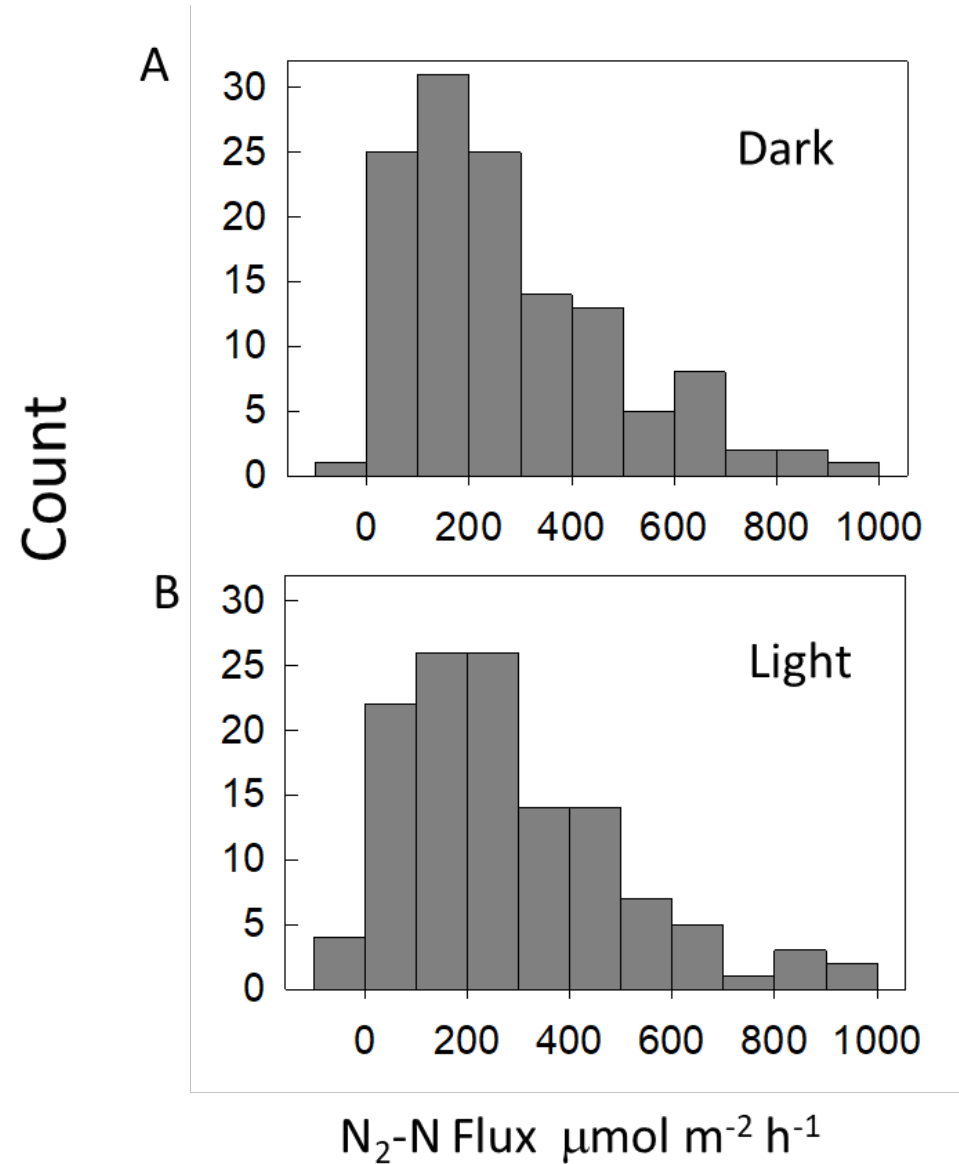
Total N Sequestered: Tissue + Shell

Enhanced Denitrification Protocol Goal:

Provide defensible, verifiable estimates of N reduction from enhanced denitrification associated with oyster reefs

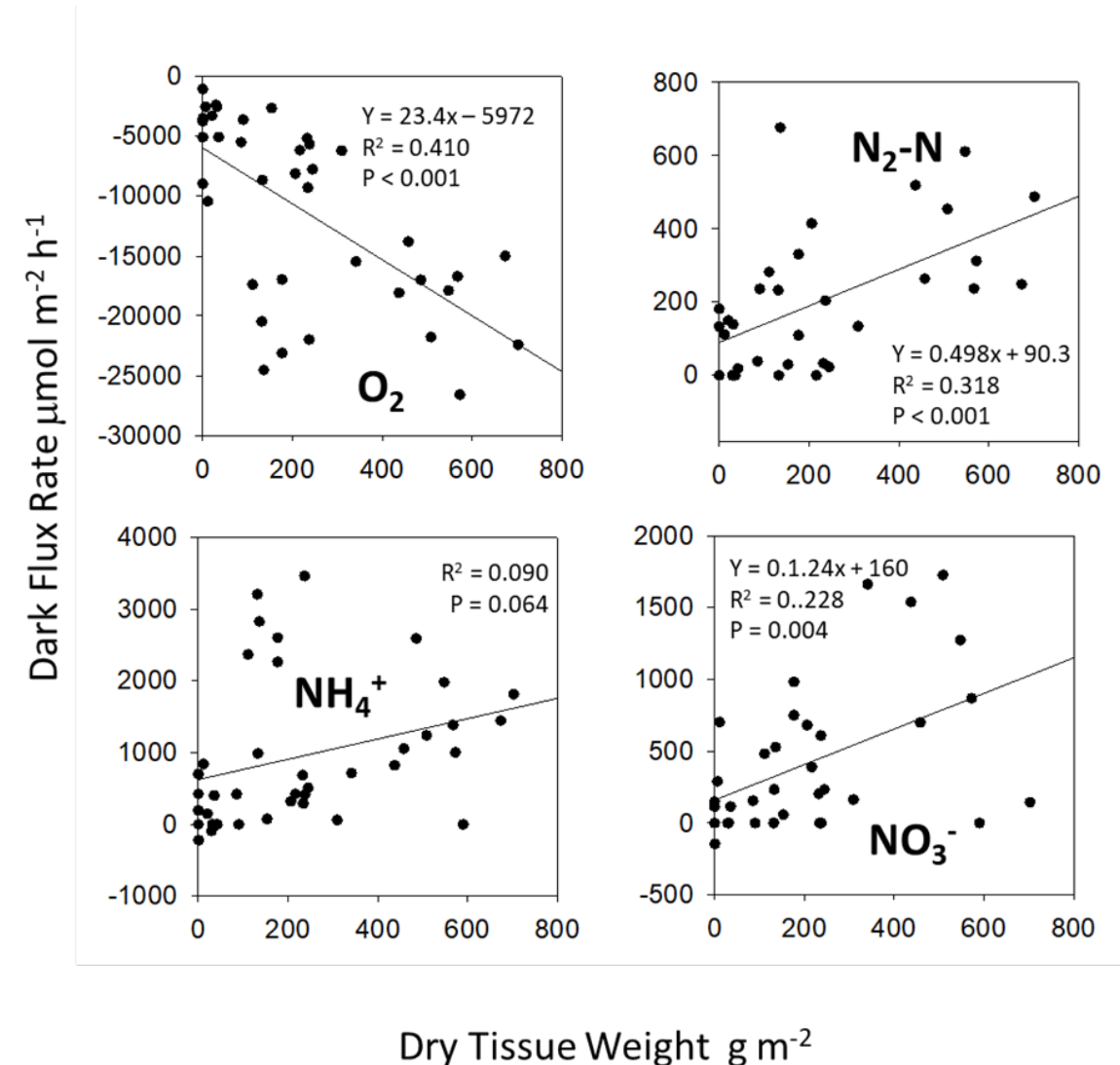


ORES Summary



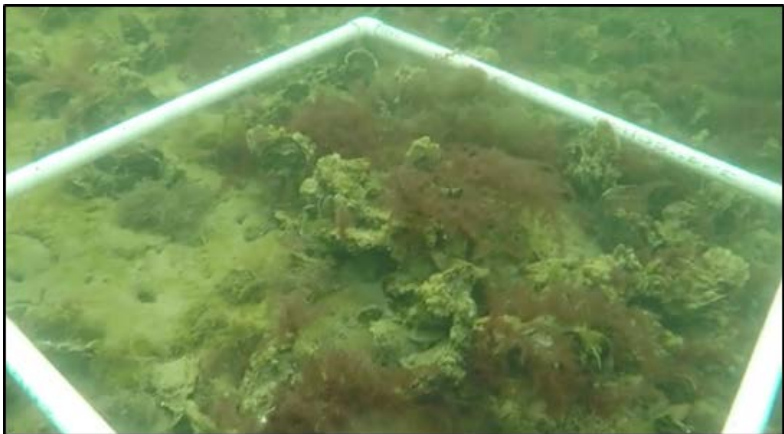
Harris Creek Observations/Conclusions

- Harris Creek rates are all “community” rates – sediments + animals
- Rates are generally high and \gg control rates
- Much of the overall nutrient processing and denitrification closely associated with oyster “clumps”
- Nutrient cycling enhanced by higher oyster biomass
- Denitrification best predicted by some non-oyster biomass – see Lisa!



Verification of Denitrification Rates using Oyster Tissue Biomass

- Research demonstrates positive relationship between denitrification rates and oyster tissue biomass
- For restoration reefs, maintenance of oyster biomass is key to maintaining the reef as a BMP



Enhanced Denitrification-Oyster Reef Restoration

Components of Site-Specific DNF Rate

- **Reference Rate (Baseline Determination)**
 - Measure denitrification (DNF) rates at oyster site and nearby reference site
 - Consult with expert to determine appropriate number of sampling points for size of BMP designated area
 - Enhanced Rate = Oyster Site – Reference Rate
- **Light and/or Dark Incubation**
 - If bottom gets sufficient light for photosynthesis ($\geq 2\%$ of incident sunlight):
 - Light incubations are needed to incorporate daylight hours for DNF rate
 - If only have dark incubations, then only dark hours can be incorporated for DNF rate
 - If no light reaches bottom then dark DNF rate can apply to full 24 hours
- **QA/QC of DNF Rate**
 - Total nitrogen (TN) fluxes should scale linearly—as O_2 decreases, TN increases

Enhanced Denitrification-Oyster Reef Restoration

Components of Site-Specific DNF Rate

- **Seasonality**

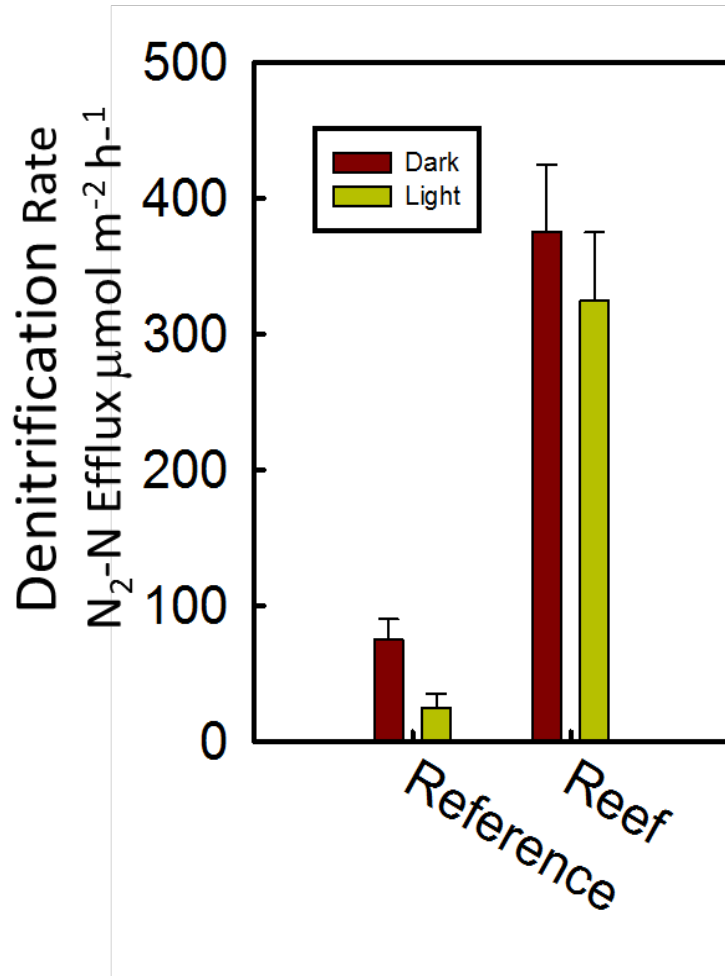
- Used defined time periods to address seasonal variability
 - Spring—Mar-May (92 days total)
 - Summer—Jun-Aug (92 days total)
 - Fall—Sep-Nov (91 days total)
 - Winter—Dec-Feb (90 days total)
- Can receive partial annual credit for seasons with measured DNF rates; full annual credit if there are DNF rates for all seasons

- **Oyster Tissue Biomass Associated with DNF Rate**

- Define “oyster presence” for verification later
 - Chamber Method: Measure oyster tissue biomass in chamber
 - Core Method: Determine average tissue biomass of designated BMP site

Enhanced Denitrification-Oyster Reef Restoration

DNF Rate Calculation Example: Light & Dark Rates (Light Reaches Bottom)



Mean Reference Rate: Dark = $60 \pm 15 \mu\text{mol N m}^{-2} \text{ h}^{-1}$
 $\pm \text{SD}$ Light = $20 \pm 5 \mu\text{mol N m}^{-2} \text{ h}^{-1}$

Mean Reef Rate: Dark = $375 \pm 75 \mu\text{mol N m}^{-2} \text{ h}^{-1}$
 $\pm \text{SD}$ Light = $325 \pm 75 \mu\text{mol N m}^{-2} \text{ h}^{-1}$

Daily rate if location and date is 14 hours light, 10 hours dark

Reference = $(60 \cdot 10 + 20 \cdot 14) = 880 \mu\text{mol m}^{-2} \text{ d}^{-1}$

Reef = $(375 \cdot 10 + 325 \cdot 14) = 8,300 \mu\text{mol m}^{-2} \text{ d}^{-1}$

Daily Enhanced Denitrification Rate:

Reef – Reference = $7,420 \mu\text{mol N m}^{-2} \text{ d}^{-1}$

= $0.10388 \text{ g N m}^{-2} \text{ d}^{-1}$

= **$0.93 \text{ lbs N acre}^{-1} \text{ d}^{-1}$**

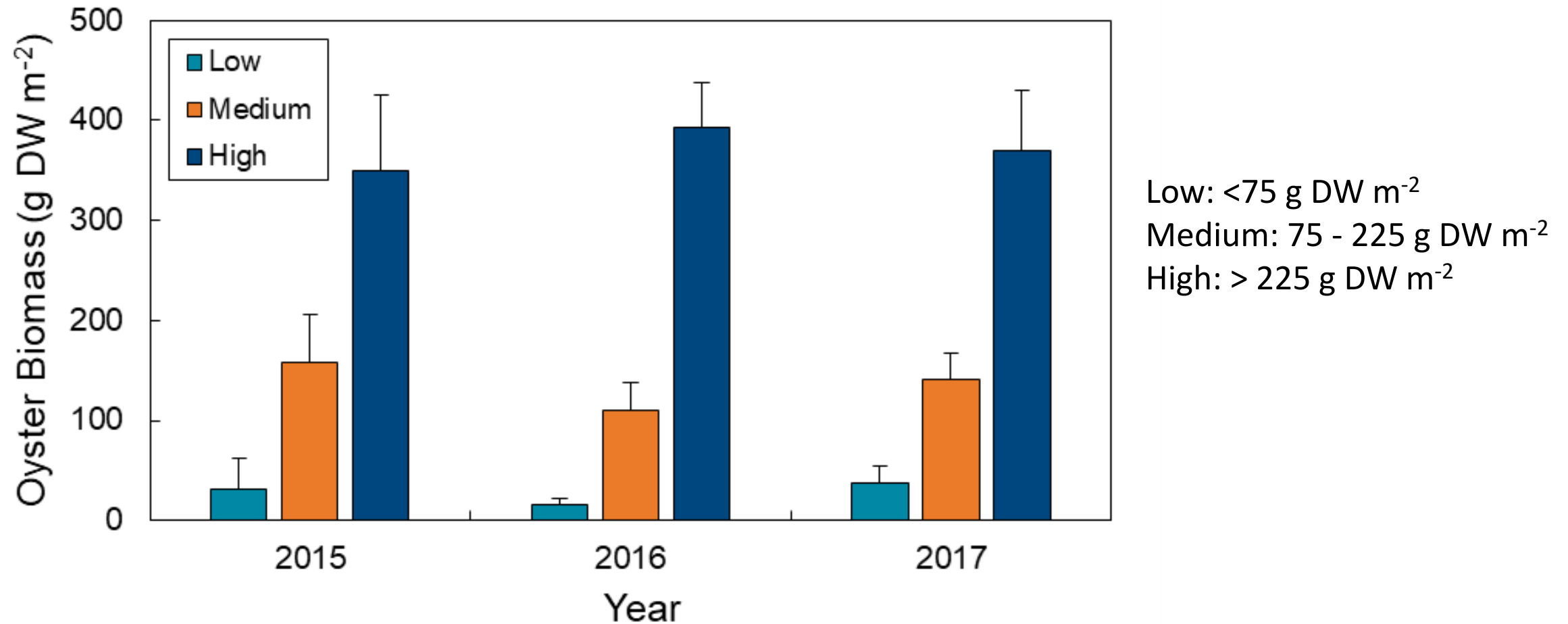
Example Assumptions:

- **Light reaches the bottom and light and dark rates assessed**
- Rates are typical warm season rates that can be observed in the summer season (e.g., July)
- Mean reference oyster tissue biomass = 0 g m^{-2}
- Mean reef oyster tissue biomass = 50 g m^{-2}

Summer Season
Jun-Aug (92 days total)

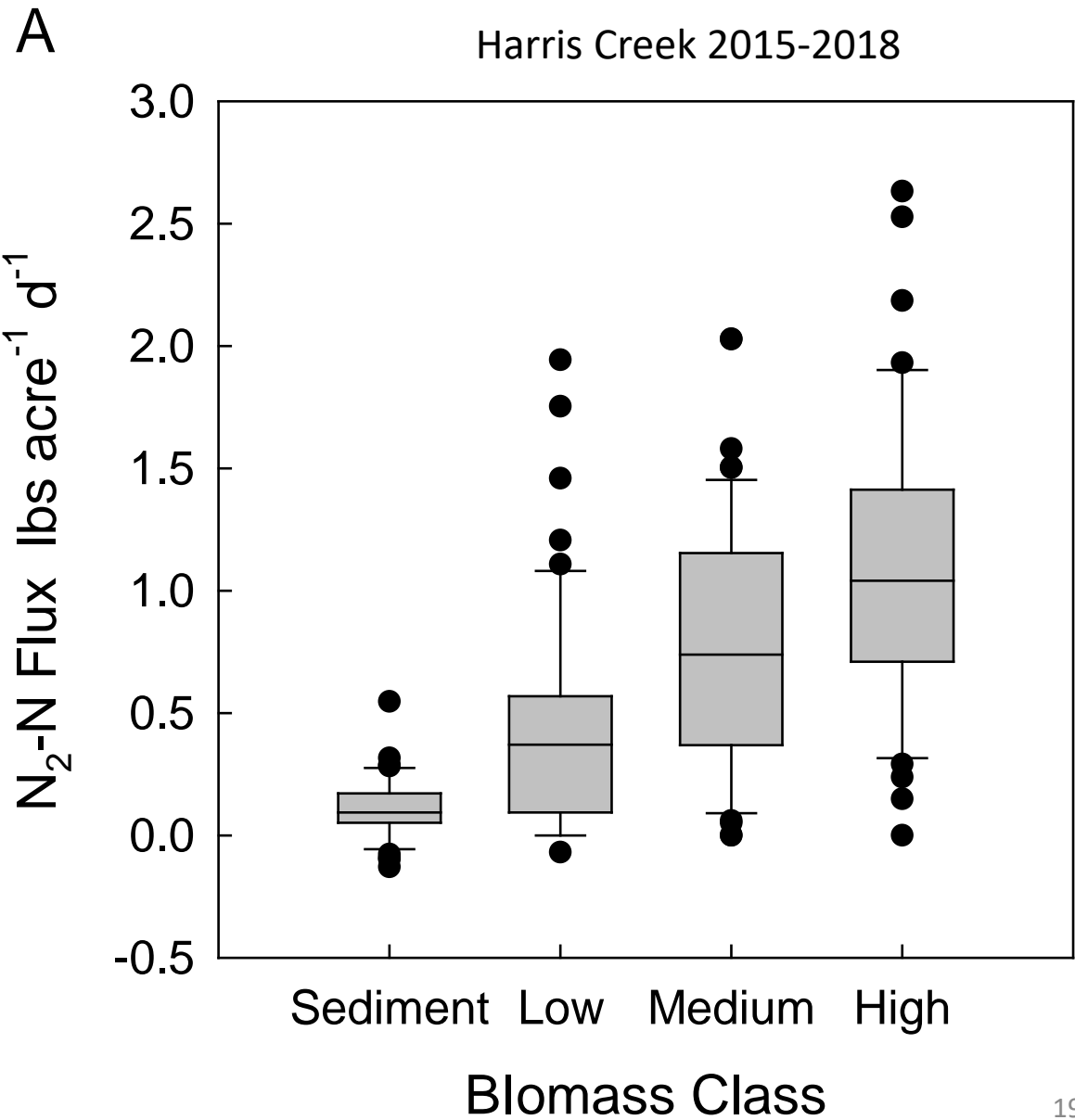
“Annual” Rate
= $0.93 \text{ lbs N acre}^{-1} \text{ d}^{-1} \cdot 92 \text{ days}$
= **$86 \text{ lbs N acre}^{-1} \text{ y}^{-1}$**

Oyster Tissue Biomass (Harris Creek, MD)



Oyster tissue biomass class definition for denitrification analysis. The data used here are from summer (June-August) tray incubations and do not include the accumulation of shell and organisms other than oysters.

Importance of Biomass



Site-Specific Best Management Practice – Oyster Reef Restoration

- Indexed to oyster biomass. Must stay relatively constant or increase to keep BMP
- BMP (*in final development*) is prescriptive, as much as possible, regarding approaches for assessment
- Implementation will not be simple, but we believe for medium to large restoration projects it could enhance the value of restoration
- Key need: sufficient oyster biomass data sets
- Key need: affordable assessment of denitrification; data beyond Harris Creek

Future Considerations – Oyster Denitrification

On-Bottom Aquaculture - Issues

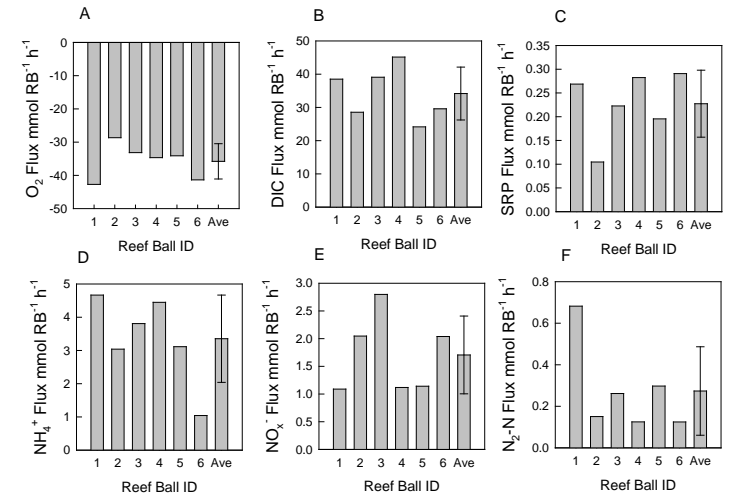
- Biomass determination
- Oysters are moved around
- Harvest effects (multiple year classes?)
- Denitrification enhancement seems likely

Water Column Aquaculture - Issues

- Poor sediment quality in footprint – sometimes
- Likely need model to explain transport/fate of biodeposits... Data to calibrate models?
- Denitrification enhancement is likely in many locations – denitrification outside of footprint

Engineered Structures

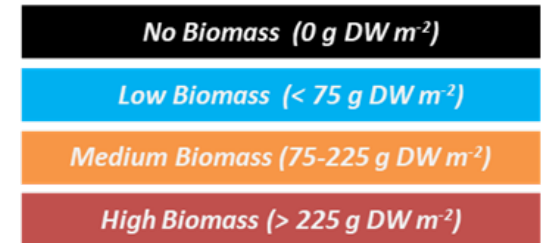
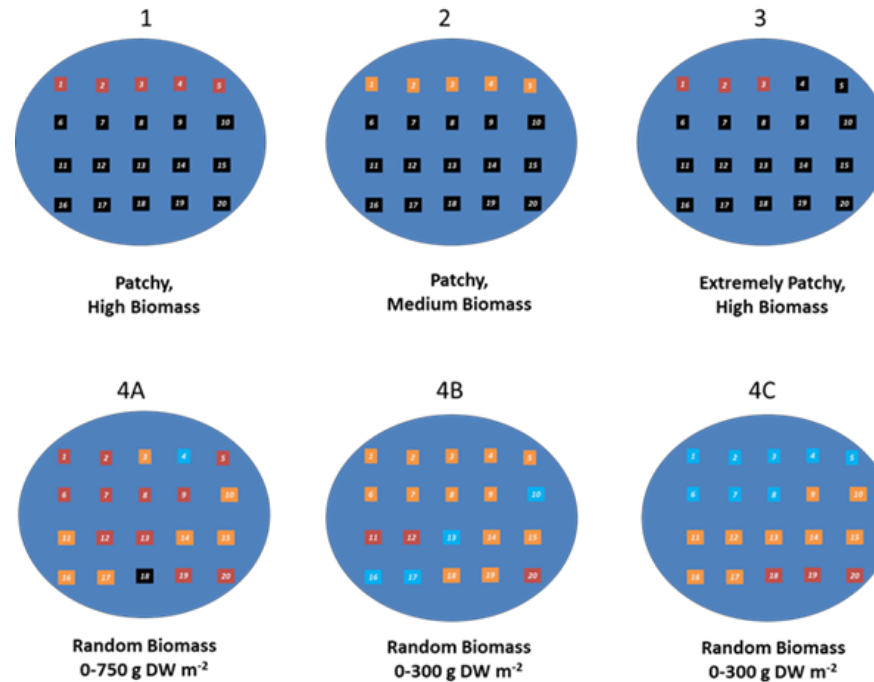
- Minimal data
- Relationship to biomass?
- Need site specific Information



Wrapping Up

Extreme patchiness could lead to whole reef estimates that could give results at variance with the “correct” amount of denitrification in a given reef. The key remaining part of this is to incorporate understandable language that ensures that common sense is applied when extrapolating data to the whole reef.

Report done in January 2020



The key challenge of this panel has been developing a plan that:

1. Captures the necessary scientific rigor required for a very new type of BMP
2. Can be implemented – agencies can develop suitable regulations and it is tractable (not excessively complicated)