# MDSG Susquehanna Flats SAV: Lyngbya field update

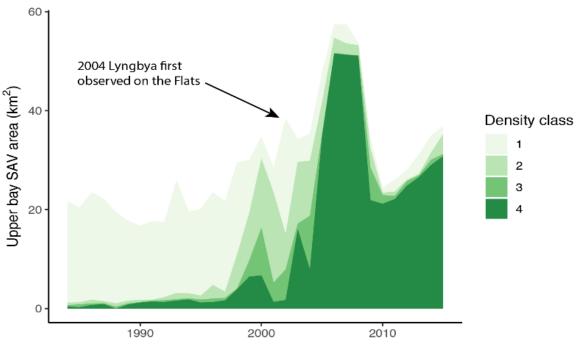


UMD: Judy O'Neil, Jeff Cornwell

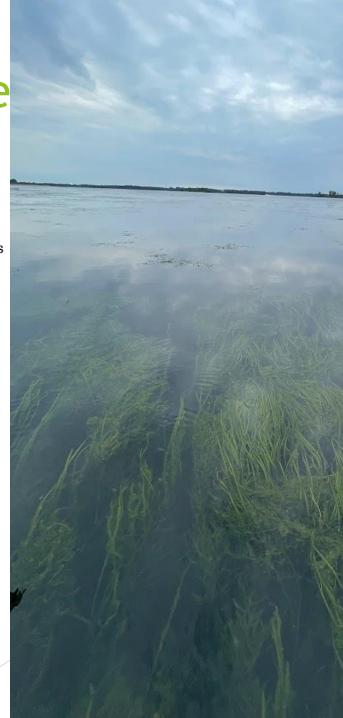
St Mary's College: Cassie Guberiz

DNR: Cathy Wazniak, Brooke Landry

## Background: SAV resurgence

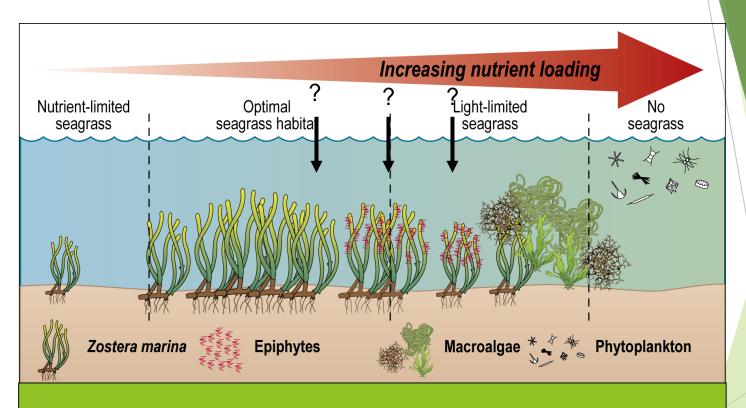


- Recovery corresponded with enhanced water clarity during extended dry period and long-term reductions in nutrient loading
- Now largest and most diverse SAV bed in Chesapeake



## Where are we in restoring SAV?

Upper Bay SAV and benthic algae increases



Are we at a tipping point between light limited and optimal seagrass conditions in the upper bay? Where high flow years push us to have higher loads and higher benthic algae??

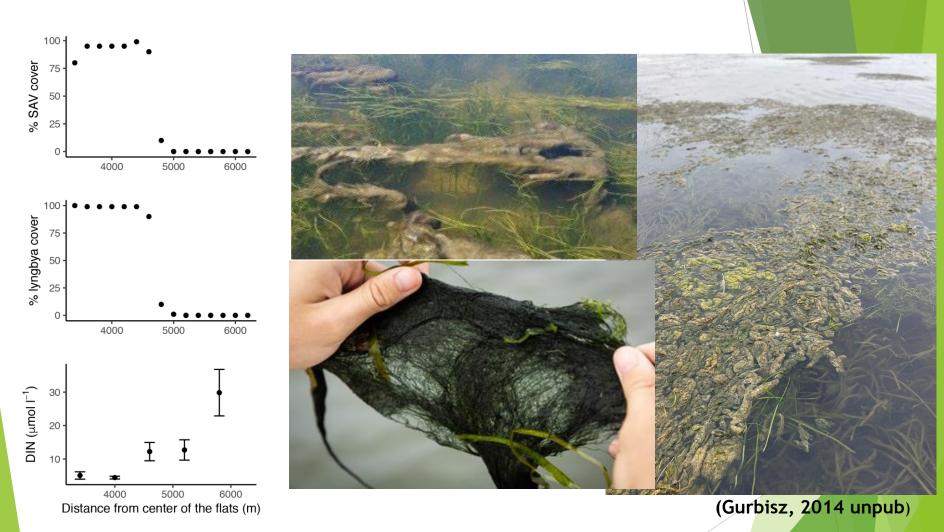
Background: Increasing benthic cyanobacteria

2004: Lyngbya latissima bloom believed to be caused by unusually high clarity in upper bay. Impacted crabbing!

- ▶ 2010: May- July, Impacting fishing gear earlier
  - Mill Creek/ furnace bay
  - ▶ July 28: algae survey, Peter Bergstrum survey
  - No microcystin toxin detected
  - Die off in September
- 2011:Winter impacts to fishing gear (Feb); poor clarity
- ▶ 2018-2020: UMD Cassie Phd work, SAV sink
  - ▶ DIN lower in center of beds
  - ▶ P low -Lyngbya likely taking it up
- ▶ Pre-2020: Watermen chlorine to clean nets/gear
- ▶ 2020: bleach shortages
  - started boiling water on vessels to clean gear



### Increases in Lyngbya, N fix?



- Data showing increased Lyngbya coverage on the inside of SAV bed;
- Low-N area in center of the SF SAV bed compared to outer edges of the bed

Why cyanobacteria in the center?



## Project Goals: Interaction of SAV and benthic cyanobacteria

- conditions that support cyanobacteria growth
- determine the effects of environmental variables (nutrients, light) on cyanobacteria production, nutrient uptake, N2 fixation, and potential toxin production
- effects of cyanobacteria on biogeochemical rate processes and SAV
- Cyanotoxins



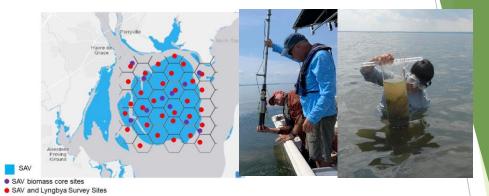




## 3 prong approach:

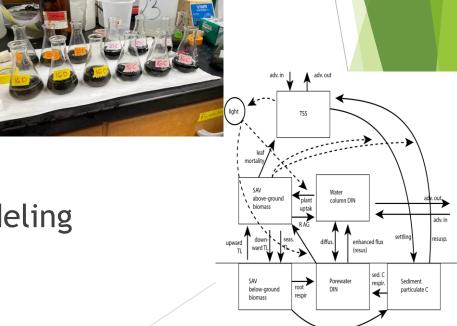


1. Field surveys

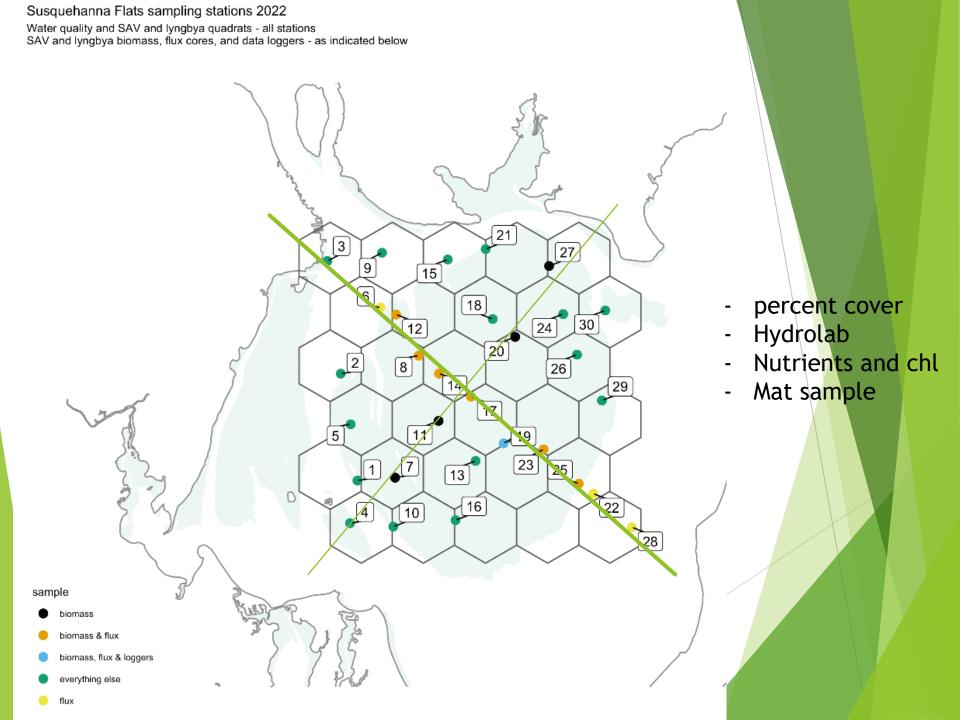


- 2. Laboratory experiments
  - bioassays
  - fluxes

3. Ecosystem simulation modeling





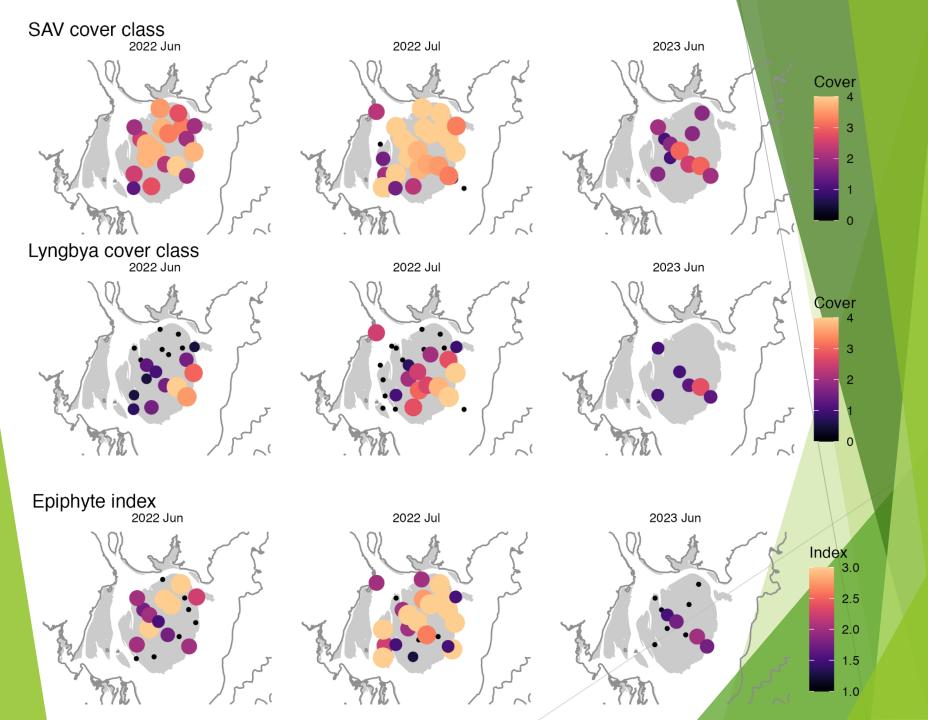


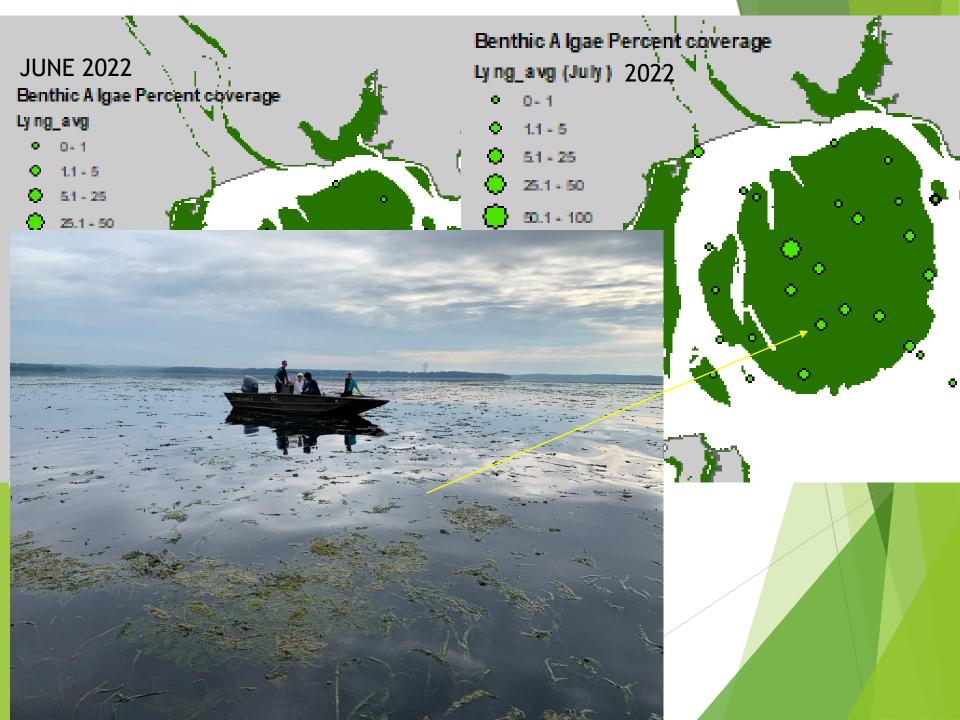
#### 2022 Methods

- Percent coverage at 30 random stations.
  - four quadrants/ site
  - Species id (SAV and benthic algae)
  - 2023 evaluating rake method from Australia
- Fluxes and Biomass Cores at transect sites.
  - Above ground: separate out the filamentous algae
    - Abundance by volumetric displacement
    - Patted dry and weighed
    - Dry weight after 24 hrs in oven
  - Below ground
    - Double washed and weighed
- Cyanotoxins on filaments
  - Greg Boyer, SUNY
  - Microcystin, cylindrospermopsin, saxitoxins
  - Filament chlorophyll a (found to be indicator of toxin in

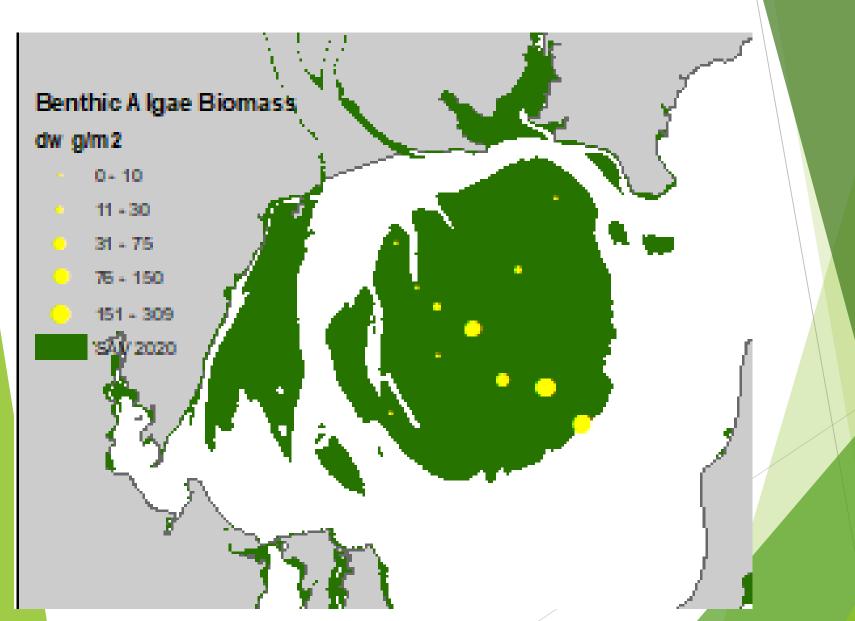








## 2022 Benthic Algae Abundance





Toxins Produced: Microcystin

fw Saxitoxins, microcystins, peptides

<sup>\*</sup>Also found green filamentous algae at several sites

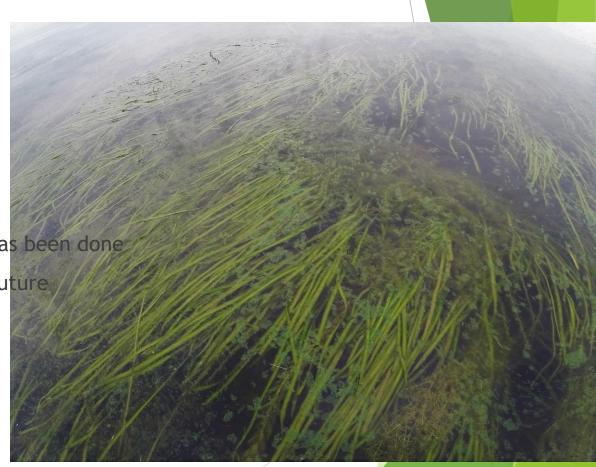






## 2022 Summary

- Missed peak cyano abundance in mid-August
  - (early june; late july)
- Water column blooms
- N fixation
- P low where Lyngbya high
- SAV biomass
- Saxitoxins detected:
  - No water toxin testing has been done
  - Samples are frozen for future



#### 2023 Field Work

- Focus on transects
- Biogeochem for models
- Rake method
- Collect water column phyto
- Bioassays
- PC:CHL fluorescence in field
- Molecular ID





# Thank you!

## Water Quality



